

## ARRAY CYTOMETRY

This application is a continuation-in-part application of Application Serial No. 09/171,550, filed on October 26, 1998, corresponding to PCT International Application No. PCT/US97/08159, filed on April 24, 1997, which in turn is based on U.S. Provisional Application Serial No. 60/016,642, filed on April 25, 1996. Applicants hereby claim the priority of these prior applications pursuant to 35 U.S.C. §§ 119 and 120. These prior applications are incorporated herein by reference.

### FIELD OF THE INVENTION

The present invention generally relates to the field of materials science and analytical chemistry.

The present invention specifically relates to the realization of a complete, functionally integrated system for the implementation of biochemical analysis in a planar, miniaturized format on the surface of a conductive and/or photoconductive substrate, with applications in pharmaceutical and agricultural drug discovery and in in-vitro or genomic diagnostics. In addition, the method and apparatus of the present invention may be used to create material surfaces exhibiting desirable topographical relief and chemical functionality, and to fabricate surface-mounted optical elements such as lens arrays.

### BACKGROUND OF THE INVENTION

I - Ions, Electric Fields and Fluid Flow: Field-induced Formation of Planar Bead Arrays

Electrokinetics refers to a class of phenomena elicited by the action of an electric field on the mobile ions surrounding charged objects in an electrolyte solution. When an object of given surface charge is immersed in a solution containing ions, a

5 diffuse ion cloud forms to screen the object's surface charge. This arrangement of a  
layer of (immobile) charges associated with an immersed object and the screening cloud  
10 of (mobile) counter-ions in solution is referred to as a "double layer". In this region of  
small but finite thickness, the fluid is not electroneutral. Consequently, electric fields  
5 acting on this region will set in motion ions in the diffuse layer, and these will in turn  
entrain the surrounding fluid. The resulting flow fields reflect the spatial distribution of  
ionic current in the fluid. Electroosmosis represents the simplest example of an  
15 electrokinetic phenomenon. It arises when an electric field is applied parallel to the  
surface of a sample container or electrode exhibiting fixed surface charges, as in the case  
20 of a silicon oxide electrode (in the range of neutral pH). As counter-ions in the  
electrode double layer are accelerated by the electric field, they drag along solvent  
molecules and set up bulk fluid flow. This effect can be very substantial in narrow  
capillaries and may be used to advantage to devise fluid pumping systems.

25 Electrophoresis is a related phenomenon which refers to the field-induced  
transport of charged particles immersed in an electrolyte. As with electroosmosis, an  
15 electric field accelerates mobile ions in the double layer of the particle. If, in contrast to  
the earlier case, the particle itself is mobile, it will compensate for this field-induced  
motion of ions (and the resulting ionic current) by moving in the opposite direction.  
30 Electrophoresis plays an important role in industrial coating processes and, along with  
electroosmosis, it is of particular interest in connection with the development of  
20 capillary electrophoresis into a mainstay of modern bioanalytical separation technology.

35 In confined geometries, such as that of a shallow experimental chamber in  
the form of a "sandwich" of two planar electrodes, the surface charge distribution and  
topography of the bounding electrode surfaces play a particularly important role in  
40 determining the nature and spatial structure of electroosmotic flow. Such a "sandwich"  
electrochemical cell may be formed by a pair of electrodes separated by a shallow gap.  
Typically, the bottom electrode will be formed by an oxide-capped silicon wafer, while  
45 the other electrode is formed by optically transparent, conducting indium tin oxide  
(ITO). The silicon (Si) wafer represents a thin slice of a single crystal of silicon which  
30 is doped to attain suitable levels of electrical conductivity and insulated from the  
electrolyte solution by a thin layer of silicon oxide (SiO<sub>x</sub>).

50 The reversible aggregation of beads into planar aggregates adjacent to an

5 electrode surface may be induced by a (DC or AC) electric field that is applied normal  
to the electrode surface. While the phenomenon has been previously observed in a cell  
formed by a pair of conductive ITO electrodes (Richetti, Prost and Barois, J. Physique  
10 Lettr. 45, L-1137 through L-1143 (1984)), the contents of which are incorporated herein  
5 by reference, it has been only recently demonstrated that the underlying attractive  
interaction between beads is mediated by electrokinetic flow (Yeh, Seul and Shraiman,  
15 "Assembly of Ordered Colloidal Aggregates by Electric Field Induced Fluid Flow",  
Nature 386, 57-59 (1997), the contents of which are incorporated herein by reference).  
This flow reflects the action of lateral non-uniformities in the spatial distribution of the  
20 current in the vicinity of the electrode. In the simplest case, such non-uniformities are  
introduced by the very presence of a colloidal bead near the electrode as a result of the  
fact that each bead interferes with the motion of ions in the electrolyte. Thus, it has  
been observed that an individual bead, when placed near the electrode surface, generates  
25 a toroidal flow of fluid centered on the bead. Spatial non-uniformities in the properties  
15 of the electrode can also be introduced deliberately by several methods to produce lateral  
fluid flow toward regions of low impedance. These methods are described in subsequent  
sections below.

30 Particles embedded in the electrokinetic flow are advected regardless of  
their specific chemical or biological nature, while simultaneously altering the flow field  
20 As a result, the electric field-induced assembly of planar aggregates and arrays applies to  
35 diverse colloidal particles including: beaded polymer resins ("beads"), lipid vesicles,  
whole chromosomes, cells and biomolecules including proteins and DNA, as well as  
metal or semiconductor colloids and clusters.

40 Important for the applications to be described is the fact that the flow-  
25 mediated attractive interaction between beads extends to distances far exceeding the  
characteristic bead dimension. Planar aggregates are formed in response to an externally  
applied electric field and disassemble when the field is removed. The strength of the  
45 applied field determines the strength of the attractive interaction that underlies the array  
assembly process and thereby selects the specific arrangement adopted by the beads  
30 within the array. That is, as a function of increasing applied voltage, beads first form  
planar aggregates in which particles are mobile and loosely packed, then assume a  
50 tighter packing, and finally exhibit a spatial arrangement in the form of a crystalline, or

5 ordered, array resembling a raft of bubbles. The sequence of transitions between states  
of increasing internal order is reversible, including complete disassembly of planar  
10 aggregates when the applied voltage is removed. In another arrangement, at low initial  
concentration, beads form small clusters which in turn assume positions within an  
5 ordered "superstructure".

## 15 II - Patterning of Silicon Oxide Electrode Surfaces

Electrode patterning in accordance with a predetermined design facilitates  
the quasi-permanent modification of the electrical impedance of the EIS (Electrolyte-  
20 Insulator-Semiconductor) structure of interest here. By spatially modulating the EIS  
impedance, electrode-patterning determines the ionic current in the vicinity of the  
electrode. Depending on the frequency of the applied electric field, beads either seek  
out, or avoid, regions of high ionic current. Spatial patterning therefore conveys  
25 explicit external control over the placement and shape of bead arrays.

15 While patterning may be achieved in many ways, two procedures offer  
particular advantages. First, UV-mediated re-growth of a thin oxide layer on a properly  
prepared silicon surface is a convenient methodology that avoids photolithographic resist  
30 patterning and etching. In the presence of oxygen, UV illumination mediates the  
conversion of exposed silicon into oxide. Specifically, the thickness of the oxide layer  
20 depends on the exposure time and may thus be spatially modulated by placing patterned  
masks into the UV illumination path. This modulation in thickness, with typical  
35 variations of approximately 10 Angstroms, translates into spatial modulations in the  
impedance of the Si/SiO<sub>x</sub> interface while leaving a flat and chemically homogeneous top  
surface exposed to the electrolyte solution. Second, spatial modulations  
40 in the distribution of the electrode surface charge may be produced by UV-mediated  
photochemical oxidation of a suitable chemical species that is first deposited as a  
25 monolayer film on the SiO<sub>x</sub> surface. This method permits fine control over local  
features of the electrode double layer and thus over the electrokinetic flow.

A variation of this photochemical modulation is the creation of lateral  
30 gradients in the EIS impedance and hence in the current generated in response to the  
applied electric field. For example, this is readily accomplished by controlling the UV  
50 exposure so as to introduce a slow lateral variation in the oxide thickness or in the

5 surface charge density. As discussed below, control over lateral gradients serves to induce lateral bead transport and facilitates the implementation of such fundamental operations as capturing and channeling of beads to a predetermined destination along  
10 conduits in the form of impedance features embedded in the Si/SiO<sub>x</sub> interface. Photo-chemical patterning of functionalized chemical overlayers also applies to other types of  
5 electrode surfaces including ITO.

### 15 III - Light-controlled Modulation of the Interfacial Impedance

The spatial and temporal modulation of the EIS-impedance in accordance  
10 with a pattern of external illumination provides the basis to control the electrokinetic forces that mediate bead aggregation. The light-modulated electrokinetic assembly of  
20 planar colloidal arrays facilitates remote interactive control over the formation, placement and rearrangement of bead arrays in response to corresponding illumination patterns and thereby offers a wide range of interactive manipulations of colloidal beads  
25 and biomolecules.

To understand the principle of this methodology, it will be helpful to  
30 briefly review pertinent photoelectric properties of semiconductors, or more specifically, those of the EIS structure formed by the Electrolyte solution (E), the Insulating SiO<sub>x</sub> layer (I) and the Semiconductor (S). The photoelectric characteristics of this structure  
20 are closely related to those of a standard Metal-Insulator-Semiconductor (MIS) or Metal-Oxide-Semiconductor (MOS) devices which are described in S.M. Sze, "The  
35 Physics of Semiconductors", 2nd Edition, Chapt. 7 (Wiley Interscience 1981), the contents of which are incorporated herein by reference.

The interface between the semiconductor and the insulating oxide layer  
40 25 deserves special attention. Crucial to the understanding of the electrical response of the MOS structure to light is the concept of a space charge region of small but finite thickness that forms at the Si/SiO<sub>x</sub> interface in the presence of a bias potential. In the  
45 case of the EIS structure, an effective bias, in the form of a junction potential, is present under all but very special conditions. The space charge region forms in response to the  
30 distortion of the semiconductor's valence and conduction bands ("band bending") in the vicinity of the interface. This condition in turn reflects the fact that, while there is a bias  
50 potential across the interface, there is ideally no charge transfer in the presence of the

insulating oxide. That is, in electrochemical language, the EIS structure eliminates Faradaic effects. Instead, charges of opposite sign accumulate on either side of the insulating oxide layer and generate a finite polarization.

In the presence of a reverse bias, the valence and conduction band edges of an n-doped semiconductor bend upward near the Si/SiO<sub>x</sub> interface and electrons flow out of the interfacial region in response to the corresponding potential gradient. As a result, a majority carrier depletion layer is formed in the vicinity of the Si/SiO<sub>x</sub> interface. Light absorption in the semiconductor provides a mechanism to create electron-hole pairs within this region. Provided that they do not instantaneously recombine, electron-hole pairs are split by the locally acting electric field, and a corresponding photocurrent flows. It is this latter effect that affords control over the electrokinetic assembly of beads in the electrolyte solution.

To understand in more detail the pertinent frequency dependence of the light-induced modulation of the EIS impedance, two aspects of the equivalent circuit representing the EIS structure are noteworthy. First, there are close analogies between the detailed electrical characteristics of the electric double layer at the electrolyte-oxide interface, and the depletion layer at the interface between the semiconductor and the insulator. As with the double layer, the depletion layer exhibits electrical characteristics similar to those of a capacitor with a voltage-dependent capacitance. As discussed, illumination serves to lower the impedance of the depletion layer. Second, given its capacitive electrical response, the oxide layer will pass current only above a characteristic ("threshold") frequency. Consequently, provided that the frequency of the applied voltage exceeds the threshold, illumination can lower the effective impedance of the entire EIS structure.

This effective reduction of the EIS impedance also depends on the light intensity which determines the rate of generation of electron-hole pairs. In the absence of significant recombination, the majority of photogenerated electrons flow out of the depletion region and contribute to the photocurrent. The remaining hole charge accumulates near the Si/SiO<sub>x</sub> interface and screens the electric field acting in the depletion region. As a result, the rate of recombination increases, and the efficiency of electron-hole separation, and hence the photocurrent, decreases. For given values of frequency and amplitude of the applied voltage, one therefore expects that as the

illumination intensity increases, the current initially increases to a maximum level and then decreases. Similarly, the impedance initially decreases to a minimum value (at maximum current) and then decreases.

This intensity dependence may be used to advantage to induce the lateral displacement of beads between fully exposed and partially masked regions of the interface. As the illumination intensity is increased, the fully exposed regions will correspond to the regions of interface of lowest impedance, and hence of highest current, and beads will be drawn into these regions. As the fully exposed regions reach the state of decreasing photocurrent, the effective EIS impedance in those regions may exceed that of partially masked regions, with a resulting inversion of the lateral gradient in current. Beads will then be drawn out of the fully exposed regions. Additionally, time-varying changes in the illumination pattern may be used to effect bead motion.

#### IV - Integration of Biochemical Analysis in a Miniaturized, Planar Format

The implementation of assays in a planar array format, particularly in the context of biomolecular screening and medical diagnostics, has the advantage of a high degree of parallelity and automation so as to realize high throughput in complex, multi-step analytical protocols. Miniaturization will result in a decrease in pertinent mixing times reflecting the small spatial scale, as well as in a reduction of requisite sample and reagent volumes as well as power requirements. The integration of biochemical analytical techniques into a miniaturized system on the surface of a planar substrate ("chip") would yield substantial improvements in the performance, and reduction in cost, of analytical and diagnostic procedures.

Within the context of DNA manipulation and analysis, initial steps have been taken in this direction (i.e., miniaturization) by combining on a glass substrate, the restriction enzyme treatment of DNA and the subsequent separation of enzyme digests by capillary electrophoresis, see, for example, Ramsey, PCT Publication No. WO 96/04547, the contents of which are incorporated herein by reference, or the amplification of DNA sequences by application of the polymerase chain reaction (PCR) with subsequent electrophoretic separation, see, for example, U.S. Patent Nos. 5,498,392 and 5,587,128 to Wilding et al., the contents of which are incorporated herein by reference.

5 While these standard laboratory processes have been demonstrated in a  
miniaturized format, they have not been used to form a complete system. A complete  
system will require additional manipulation such as front-end sample processing, binding  
10 and functional assays and the detection of small signals followed by information  
5 processing. The true challenge is that of complete functional integration because it is  
here that system architecture and design constraints on individual components will  
manifest themselves. For example, a fluidic process is required to concatenate analytical  
15 steps that require the spatial separation, and subsequent transport to new locations, of  
sets of analyte. Several possibilities have been considered including electroosmotic  
20 pumping and transport of droplets by temperature-induced gradients in local surface  
tension. While feasible in demonstration experiments, these techniques place rather  
severe requirements on the overall systems lay-out to handle the very considerable DC  
voltages required for efficient electroosmotic mixing or to restrict substrate heating when  
25 generating thermally generated surface tension gradients so as to avoid adverse effects  
15 on protein and other samples.

#### Summary of the Invention

30 The present invention combines three separate functional elements to  
provide a method and apparatus facilitating the real-time, interactive spatial manipulation  
20 of colloidal particles ("beads") and molecules at an interface between a light sensitive  
electrode and an electrolyte solution. The three functional elements are: the electric  
35 field-induced assembly of planar particle arrays at an interface between an insulating or  
a conductive electrode and an electrolyte solution; the spatial modulation of the  
interfacial impedance by means of UV-mediated oxide regrowth or surface- chemical  
40 patterning; and, finally, the real-time, interactive control over the state of the interfacial  
impedance by light. The capabilities of the present invention originate in the fact that  
25 the spatial distribution of ionic currents, and thus the fluid flow mediating the array  
assembly, may be adjusted by external intervention. Of particular interest is the  
45 introduction of spatial non-uniformities in the properties of the pertinent EIS structure.  
As described herein, such inhomogeneities, either permanent or temporary in nature,  
30 may be produced by taking advantage of the physical and chemical properties of the EIS  
50 structure.



5 The invention relates to the realization of a complete, functionally  
integrated system for the implementation of biochemical analysis in a planar,  
miniaturized format on the surface of a silicon wafer or similar substrate. In addition,  
10 the method and apparatus of the present invention may be used to create material  
5 surfaces exhibiting desirable topographical relief and chemical functionality, and to  
fabricate surface-mounted optical elements such as lens arrays.

15 The combination of three functional elements endows the present  
invention with a set of operational capabilities to manipulate beads and bead arrays in a  
planar geometry to allow the implementation of biochemical analytical techniques.  
20 These fundamental operations apply to aggregates and arrays of colloidal particles  
including: beaded polymer resins also referred to as latices, vesicles, whole chromo-  
somes, cells and biomolecules including proteins and DNA, as well as metal or semicon-  
ductor colloids and clusters.

25 Sets of colloidal particles may be captured, and arrays may be formed in  
15 designated areas on the electrode surface (Figs. 1a, 1b and Figs. 2a-d). Particles, and  
the arrays they form in response to the applied field, may be channeled along conduits of  
any configuration that are either embedded in the Si/SiO<sub>x</sub> interface by UV-oxide  
30 patterning or delineated by an external pattern of illumination. This channeling (Figs.  
1c, 1d, 1e, Figs. 3c, 3d), in a direction normal to that of the applied electric field, relies  
20 on lateral gradients in the impedance of the EIS structure and hence in the field-induced  
current. As discussed herein, such gradients may be introduced by appropriate patterns  
of illumination, and this provides the means to implement a gated version of  
35 translocation (Fig. 1e). The electrokinetic flow mediating the array assembly process  
may also be exploited for the alignment of elongated particles, such as DNA, near the  
40 surface of the electrode. In addition, the present invention permits the realization of  
25 methods to sort and separate particles.

45 Arrays of colloidal particles may be placed in designated areas and  
confined there until released or disassembled. The overall shape of the array may be  
delineated by UV-oxide patterning or, in real time, by shaping the pattern of  
30 illumination. This capability enables the definition of functionally distinct compart-  
ments, permanent or temporary, on the electrode surface. Arrays may be subjected to  
50 changes of shape imposed in real time, and they may be merged with other arrays (Fig.

5 1f) or split into two or more subarrays or clusters (Fig. 1g, Figs. 4a, 4b). In addition,  
the local state of order of the array as well as the lateral particle density may be  
reversibly adjusted by way of the external electric field or modified by addition of a  
10 second, chemically inert bead component.

5 The present invention also allows for the combination of fundamental  
operations to develop increasingly complex products and processes. Examples given  
herein describe the implementation of analytical procedures essential to a wide range of  
15 problems in materials science, pharmaceutical drug discovery, genomic mapping and  
sequencing technology. Important to the integration of these and other functionalities in  
20 a planar geometry is the capability, provided by the present invention, to impose tempo-  
rary or permanent compartmentalization in order to spatially isolate concurrent processes  
or sequential steps in a protocol and the ability to manipulate sets of particles in a  
manner permitting the concatenation of analytical procedures that are performed in  
25 different designated areas on the substrate surfaces.

15 This invention is for a method and apparatus to direct the lateral motion  
and induce the assembly into planar arrays of cells on semiconductor surfaces in  
response to temporally and spatially varying electric fields and to projected patterns of  
30 illumination.

The response of different types of cells to variations in the frequency and  
20 voltage of the applied electric field and to specific patterns and changing intensity of  
illumination enables the on-cue assembly of multiple arrays of cells in any desired  
35 position on the substrate, and further enables the repositioning, disassembly and re-  
assembly and more generally the reconfiguration and segmentation of such arrays. Time-  
constant and time-varying illumination patterns are generated by an optically  
40 programmable illumination pattern generator may be used and can be based on a  
graphical representation of the pattern created on a personal computer. The  
novel methodology of image cytometry introduced in the present invention relates to the  
45 general fields of molecular and cellular biology and has applications in the fields of  
diagnostics, genetic analysis, cell biology and drug discovery. The image cytometry  
30 facilitates a multiplicity of cell-based biological and biochemical procedures based on the  
quantitative analysis of images recorded from array of cells. As with Random Encoded  
50 Array Detection (READ) or Programmable Array Reconfiguration and Segmentation

(PARSE), these assays are performed in a highly parallel format in a small volume of sample on the surface of a semiconductor wafer.

#### Brief Description of Drawings

Other objects, features and advantages of the invention discussed in the above brief explanation will be more clearly understood when taken together with the following detailed description of an embodiment which will be understood as being illustrative only, and the accompanying drawings reflecting aspects of that embodiment, in which:

Figs. 1a-h are illustrations of the fundamental operations for bead manipulation;

Figs. 2a and 2b are photographs illustrating the process of capturing particles in designated areas on the substrate surface;

Figs. 2c and 2d are photographs illustrating the process of excluding particles from designated areas on the substrate surface;

Figs. 3a and 3b are illustrations of the oxide profile of an Si/SiO<sub>x</sub> electrode;

Figs. 3c and 3d are photographs of the channeling of particles along conduits;

Figs. 4a and 4b are photographs of the splitting of an existing aggregate into small clusters;

Fig. 5 is a photograph of the lensing action of individual colloidal beads;

Figs. 6a-c are side view illustrations of a layout-preserving transfer process from a microtiter plate to a planar cell;

Fig. 7 is a photograph of the inclusion of spacer particles within bead clusters;

Fig. 8 is an illustration of binding assay variations;

Figs. 9a and 9b are illustrations of two mechanisms of particle sorting, and Fig. 9c is an illustration of the oxide profile of Fig. 9b;

Fig. 10 is an illustration of a planar array of bead-anchored probe-target complexes;

Figs. 11a-11e are illustrations of DNA stretching in accordance with the

present invention;

Fig. 12 is an illustration of *E.coli* cells (grown to log phase and suspended in 280 mM mannitol solution at the density of  $10^8$ /ml) which form arrays under electric field. (Bright field image. 25x obj, NA 0.45. Electric field: 5V, 200 Hz);

Fig. 13 is an illustration of the same effect as in Figure 12, with *S. cerevisiae* cells (baker's yeast). (Bright field image. 25x obj, NA 0.45; electric field: 5V, 400 Hz);

Fig. 14 is an illustration of *E.coli* forming diamond shapes that are projected to the surface by an LCD panel controlled by appropriate graphics software; (20x objective, NA 0.28; Electric field: 10V, 7 kHz);

Fig. 15. is an illustration of *S. cerevisiae* responding to two circular illumination patterns; (20x objective, NA 0.28; Electric field: 15 V, 15 kHz)

Fig. 16 is an illustration of *S. cerevisiae* responding to illumination, with an arrow indicate the direction cells are moving in or being expelled from the illuminated regions (20x objective, NA 0.28; Electric field: 15 V, 15 kHz);

Fig. 17 is an illustration of *E. coli* cells being repelled from the illumination circle (20x objective, NA 0.28; Electric field: 10 V, 7 kHz; low substrate doping level (1 Ohm cm-10 Ohm cm));

Fig. 18 is an illustration of *E.coli* cells following the expansion of the rectangular illuminated shape (20x objective, NA 0.28; Electric field: 15 V, 8 kHz);

Fig. 19 is an illustration of *E. coli* cells being trapped by the light stripe at the left and being prevented from moving to the circular region on the right (20x objective, NA 0.28; Electric field: 15 V, 8 kHz);

Fig. 20 is an illustration of an illuminated region containing both types of cells being suddenly expanded or "dragged", with *E. coli* cells (left) responding faster than the larger *S. cerevisiae* cells (right), resulting in segregation. (Bright field image. 25x obj, NA 0.45 Electric field: 5V, 400 Hz);

Fig. 21 is an illustration of immunophenotyping according to the present invention;

Fig. 22 is an illustration of affinity cell detection according to the present invention;

Fig. 23 is an illustration of cell sorting and isolation according to the

present invention;

Fig. 24 is an illustration of cell-based screening according to the present invention;

Fig. 25 is an illustration of cell-secretion based screening according to the present invention;

Fig. 26 is an illustration of cell to cell interaction according to the present invention; and

Fig. 27 is an illustration of chemotaxis analysis according to the present invention.

#### Detailed Description of the Preferred Embodiments

The three functional elements of the present invention may be combined so as to provide a set of fundamental operations for the interactive spatial manipulation of colloidal particles and molecules, assembled into planar aggregates adjacent to an electrode surface. In the following description, fundamental operations in this "toolset" are described in order of increasing complexity. Specifically, it is useful to adopt a classification scheme based on the total number of inputs and outputs, or "terminals", involved in a given operation. For example, the merging of two separate arrays, or sets of particles, into one would be a "three-terminal" operation, involving two inputs and one output. The converse three-terminal operation, involving one input and two outputs, is the splitting of a given array into two subarrays.

Experimental conditions yielding the phenomena depicted in the various photographs included herein are as follows. An electrochemical cell is formed by a pair of planar ITO electrodes, composed of an ITO layer deposited on a glass substrate, or by a Si/SiO<sub>x</sub> electrode on the bottom and an ITO electrode on the top, separated by a typical gap of 50 microns or less. Given its dependence on the photoelectric properties of the Si/SiO<sub>x</sub> interface, light control is predicated on the use of a Si/SiO<sub>x</sub> electrode. Leads, in the form of platinum wires, are attached to the ITO and to the silicon electrode, which is first etched to remove the insulating oxide in the contact region, by means of silver epoxy. The cell is first assembled and then filled, relying on capillary action, with a suspension of colloidal beads, 1 or 2 microns in diameter, at a typical

5 concentration of 0.1% solids in 0.1mM azide solution, corresponding to approximately  
2x10<sup>8</sup> particles per milliliter. The number is chosen so as to yield between ½ and 1  
10 full monolayer of particles on the electrode surface. Anionic (e.g., carboxylated  
polystyrene, silica), cationic (e.g., aminated polystyrene) or nominally neutral (e.g.,  
5 polystyrene) have all been used to demonstrate the basic phenomena underlying the three  
functional elements of the present invention. The silicon electrode was fabricated from a  
15 1 inch-square portion of a Si (100) wafer, typically 200-250 microns thick, n-doped to  
typically 0.01 Ohm cm resistivity, and capped with a thin oxide of typically 30-40  
Angstroms thickness. A thick oxide layer of typically 6000-8000 Angstrom thickness,  
20 grown under standard conditions in a furnace at 950 degrees C, may be etched by  
standard photolithography to define the structures of interest. Alternatively, a thin oxide  
layer may be regrown on a previously stripped surface of (100)-orientation under UV  
illumination. Given its ease of implementation and execution, UV-mediated oxide  
25 regrowth is the preferable technique: it provides the means to pattern the surface by  
placing a quartz mask representing the desired pattern in the path of UV illumination and  
it leaves a chemically homogeneous, topographically flat top surface. To avoid  
30 non-specific particle adsorption to the electrode surface, stringent conditions of  
cleanliness should be followed, such as those set forth in the General Experimental  
Conditions below.

35 The fundamental one-terminal operation is a "capture-and-hold" operation  
(Fig. 1a) which forms an array of particles in a designated area of arbitrary outline on  
the surface that is delineated by UV-mediated oxide patterning or by a corresponding  
pattern of illumination projected on an otherwise uniform Si/SiOx substrate surface.  
40 Figs. 2a and 2b illustrate bead capture on a surface characterized by a very thin oxide  
region 22 (approximately 20-30 Angstroms in thickness) and correspondingly low  
impedance, while the remaining surface is covered with the original, thick oxide with  
correspondingly high impedance. In Fig. 2a, there is no applied field, and hence, no  
45 bead capture. In contrast, in Fig. 2b, an electric field is applied (10Vp-p source, 1 kHz)  
and bead capture occurs within the thin oxide region 22. Under these conditions, an  
30 array starts to grow within less than a second and continues to grow over the next  
approximately 10 seconds as beads arrive from increasingly larger distances to add to  
the outward growing perimeter of region 22. Growth stops when the array approaches

5 the outer limit of the delineated target area, i.e., the area defined by the thin oxide  
having a low impedance. The internal state of order of the captured aggregate of beads  
is determined by the strength of the  
10 applied voltage, higher values favoring increasingly denser packing of beads and the  
5 eventual formation of ordered arrays displaying a hexagonally crystalline configuration  
in the form of a bubble raft. The array remains in place as long as the applied voltage is  
present. Removal of the applied voltage results in the disassembly of the array.

15 The "capture-and-hold" operation may also be implemented under  
illumination with visible or infrared light, for example by simply projecting a mask  
20 patterned with the desired layout onto the Si/SiO<sub>x</sub> electrode. A regular 100W quartz  
microscope illuminator has been used for this purpose on a Zeiss UEM microscope, with  
apertures or masks inserted in the intermediate image plane to provide the required  
shape in the plane of the electrode (when focused properly under conditions of Koehler  
illumination). Alternatively, an IR laser diode with output of 3 mW at 650 - 680nm also  
25 has been used. The use of external illumination rather than oxide patterning for the  
spatial confinement of particles allows the confinement pattern to be easily modified.

30 Related to "capture-and-hold" is the one-terminal operation of "exclude-  
and-hold" (Fig. 1b) which clears particles from a designated area on the surface.  
Increasing the frequency of the applied voltage to approximately 100kHz leads to an  
20 inversion in the preference of particles which assemble in the thin-oxide portion of the  
surface (e.g., region 22, Fig. 2b) and instead form structures decorating the outside of  
the target area perimeter. Rather than relying on this effect, the exclusion of particles  
from the desired areas is also accomplished, in analogy to the original  
"capture-and-hold" operations, by simply embedding the corresponding structure in the  
40 Si/SiO<sub>x</sub> interface by UV-mediated oxide regrowth. In the example of Figs. 2c and 2d,  
this is achieved, under conditions otherwise identical to those described above, with  
respect to Figs. 2a and 2b, by applying 20V (pp) at 10kHz. While the oxide thickness in  
the non designated areas 24 is approximately 30 Angstroms, the value in the designated  
square areas 26 is approximately 40 Angstroms, implying a correspondingly higher  
45 impedance at the applied frequency.

50 The "capture-and-hold" operation enables the spatial  
compartmentalization of the substrate surface into functionally distinct regions. For

5 example, particles of distinct chemical type, introduced into the electrochemical cell at different times or injected in different locations, can be kept in spatially isolated locations by utilizing this operation.

10 The fundamental two-terminal operation is translocation (Fig. 1c), or the controlled transport of a set of particles from location O to location F on the surface; here, O and F are target areas to which the above-described one-terminal operations may be applied. The one-dimensional, lateral bead transport used in translocation is achieved by imposing a lateral current along a conduit connecting areas O and F, as shown in 15 Figs. 3a and 3b or by projecting a corresponding linear pattern of illumination. In this channeling operation, beads move in the direction of lower impedance in the direction of the arrow shown in Figs. 3a and 3b, in accordance with the underlying electrokinetic flow.

Oxide patterning may be utilized in two ways to create a lateral current along the Si/SiO<sub>x</sub> interface. The simplest method is depicted in Fig. 3c and shows a 25 large open holding area 32 fed by three narrow conduits 34 defined by etching a thermal oxide. Beads move to the holding area 32 along the narrow conduits 34 to form a bead array. Fig. 3d is a large scale view of the array of Fig. 3c. The principle invoked in creating transport is that of changing the aspect ratio (narrow conduit connected to wide holding area) of the embedded pattern with constant values of thin oxide thickness inside 30 and thick oxide outside, as illustrated in Fig. 3a. In Figs. 3c and 3d, the applied voltage was 10V (pp) at 10kHz. An alternative approach for creating bead transport, enabled by UV-mediated oxide regrowth, is to vary the oxide thickness along the conduit in a controlled fashion. This is readily accomplished by UV exposure through a graduated filter. Differences in the oxide thickness between O and F of as little as 5-10 Angstroms 35 suffice to effect lateral transport. In this situation, the aspect ratio of the conduit and holding areas need not be altered. This is illustrated in Fig. 3b.

The use of external illumination to define conduits, by varying the 45 illumination intensity along the conduit to create the requisite impedance gradient, has the advantage that the conduit is only a temporary structure, and that the direction of motion may be modified or reversed if so desired. The present invention provides for 30 mechanisms of light-mediated active linear transport of planar aggregates of beads under interactive control. This is achieved by adjusting an external pattern of illumination in



5 real time, either by moving the pattern across the substrate surface in such a way as to  
entrain the illuminated bead array or by electronically modulating the shape of the  
pattern to induce motion of particles.

10 Two modes of light-mediated, active transport are:

- 5
- i) Direct Translocation ("tractor beam") which is a method of translocating arrays  
15 and of delineating their overall shape by adjusting parameters so as to favor particle  
assembly within illuminated areas of the surface, as described herein. Arrays simply  
follow the imposed pattern. The rate of motion is limited by the mobility of particles in  
20 the fluid and thus depends on particle diameter and fluid viscosity.
- ii) Transverse Array Constriction is a bead transport mechanism  
related to peristaltic pumping of fluids through flexible tubing. The light-control  
25 component of the present invention may be used for a simple implementation of this very  
general concept. A multi-component planar aggregate of beads is confined to a  
15 rectangular channel, by UV-patterning if so desired, or simply by light. Beads are free  
to move along the channel by diffusion (in either direction). An illumination pattern  
30 matching the transverse channel dimension is set up and is then varied in time so as to  
produce a transverse constriction wave that travels in one direction along the channel.  
Such a constriction wave may be set up in several ways. A conceptually simple method  
20 is to project a constricting mask onto the sample and move the projected mask pattern in  
the desired fashion. This method also may be implemented electronically by controlling  
35 the illumination pattern of a suitable array of light sources, thus obviating the need for  
moving parts in the optical train.
- The control of lateral bead transport by changing or moving patterns of  
40 illumination has the advantage that it may be applied whenever and wherever (on a given  
substrate surface) required, without the need to impose gradients in impedance by  
predefined UV patterning. On the other hand, a predefined impedance pattern can  
45 provide additional capabilities in conjunction with light-control. For example, it may be  
desirable to transport beads against a substrate-embedded impedance gradient to separate  
30 beads on the basis of mobility.

50 Conduits connecting O and F need not be straight: as with tracks directing  
the motion of trains, conduits may be shaped in any desirable fashion (Fig. 1d). A gated

5 version of translocation (Fig. 1e) permits the transport of particles from O to F only after the conduit is opened (or formed in real time) by a gating signal. This operation utilizes UV oxide patterning to establish two holding areas, O and F, and also light  
10 control to temporarily establish a conduit connecting O and F. An alternative implementation is based on an oxide embedded impedance gradient. A zone along the conduit is illuminated with sufficiently high intensity to keep out particles, thereby blocking the passage. Removal (or reduction in intensity) of the illumination opens the  
15 conduit. In the former case, light enables the transport of beads, while in the latter case, light prevents the transport of beads.

20 The fundamental three-terminal operations are the merging and splitting of sets or arrays of beads (Figs. 1f and 1g). The merging of two arrays (Fig. 1f) involves the previous two fundamental operations of "capture-and-hold", applied to two spatially isolated sets of beads in locations O1 and O2, and their respective channeling along merging conduits into a common target area, and their eventual channeling, subse-  
25 quent to mixing, or a chemical reaction, into the final destination, a third holding area, F. This is accomplished, under the conditions stated above, by invoking one-terminal and gated two-terminal operations.

30 The splitting of an array into two subarrays (Fig. 1g) is a special case of a generally more complex sorting operation. Sorting involves the classification of beads in a given set or array into one of two subsets, for example according to their fluores-  
35 cence intensity. In the simpler special case, a given array, held in area O, is to be split into two subarrays along a demarcation line, and subarrays are to be moved to target areas F1 and F2. Under the conditions stated above, this is accomplished by applying the "capture-and-hold" operation to the array in O. Conduits connect O to F1 and F2.  
40 High intensity illumination along a narrowly focused line serves to divide the array in a defined fashion, again relying on gated translocation to control transport along conduits away from the holding area O. An even simpler version, termed indiscriminate  
45 splitting, randomly assigns particles into F1 and F2 by gated translocation of the array in O into F1 and F2 after conduits are opened as described above.

50 Figs. 4a and 4b show a variant in which beads in region O (Fig. 4a) are split into multiple regions F1, F2, ... Fn (Fig. 4b). This reversible splitting of an aggregate or array into n subarrays, or clusters, is accomplished, for carboxylated

5 polystyrene spheres of 2 micron diameter at a concentration corresponding to an  
electrode coverage of a small fraction of a monolayer, at a frequency of 500Hz, by  
10 raising the applied voltage from typically 5V (pp) to 20V (pp). This fragmentation of an  
array into smaller clusters reflects the effect of a field-induced particle polarization. The  
5 splitting is useful to distribute particles in an array over a wider area of substrate for  
presentation to possible analytes in solution, and for subsequent scanning of the  
15 individual clusters with analytical instruments to make individual readings.

The three functional elements of the present invention described herein  
may be also combined to yield additional fundamental operations to control the orien-  
10 tation of anisotropic objects embedded in the electroosmotic flow created by the applied  
20 electric field at the electrode surface. The direction of the flow, in the plane of the  
substrate, is controlled by gradients in the impedance that are shaped in the manner  
described in connection with the channeling operation. This is used to controllably align  
25 anisotropic objects as illustrated in Fig. 1h, and may be applied to stretch out and align  
15 biomolecules, such as DNA.

An additional fundamental operation that complements the previous set is  
that of permanently anchoring an array to the substrate. This is best accomplished by  
30 invoking anchoring chemistries analogous to those relying on heterobifunctional  
cross-linking agents invoked to anchor proteins via amide bond formation. Molecular  
20 recognition, for example between biotinylated particles and surface-anchored  
35 streptavidin, provides another class of coupling chemistries for permanent anchoring.

#### General Experimental Conditions

40 The functional elements, namely the electric-field induced assembly of  
25 planar particle arrays, the spatial modulation of the interfacial impedance by means of  
UV-mediated oxide or surface-chemical patterning and finally, the control over the state  
of the interfacial impedance by light which are used in the present invention, have been  
45 demonstrated in experimental studies. These studies employed n-doped silicon wafers  
(resistivities in the range of 0.01 Ohm cm), capped with either thermally grown oxide  
30 layers of several thousand Angstrom thickness, or with thin oxide layers, regrown after  
removal of the original "native" oxide in HF, under UV illumination from a deuterium  
50 source in the presence of oxygen to typical thicknesses between 10 and 50 Angstroms.

5 Lithographic patterning of thermally grown oxide employed standard procedures implemented on a bench top (rather than a clean room) to produce features in the range of several microns.

10 Surfaces were carefully cleaned in adherence with industry standard RCA and Piranha cleaning protocols. Substrates were stored in water produced by a Millipore water purification system prior to use. Surfaces were characterized by measuring the contact angle exhibited by a 20 microliter droplet of water placed on the surface and viewed (from the side) through a telescope. The contact angle is defined as the angle subtended by the surface and the tangent to the droplet contour (in side view) at the point of contact with the surface. For example, a perfectly hemispherical droplet shape would correspond to a contact angle of 90 degrees. Surface chemical derivatization with mercapto-propyl-trimethoxysilane (2% in dry toluene) produced surfaces giving typical contact angles of 70 degrees. Oxidation of the terminal thiol functionality under UV irradiation in the presence of oxygen reduced the contact angle to zero in less than 10 min of exposure to UV from the deuterium source. Other silane reagents were used in a similar manner to produce hydrophobic surfaces, characterized by contact angles in excess of 110 degrees.

30 Simple "sandwich" electrochemical cells were constructed by employing kapton film as a spacer between Si/SiO<sub>x</sub> and conductive indium tin oxide (ITO), deposited on a thin glass substrate. Contacts to platinum leads were made with silver epoxy directly to the top of the ITO electrode and to the (oxide-stripped) backside of the Si electrode. In this two-electrode configuration, AC fields were produced by a function generator, with applied voltages ranging up to 20V and frequencies varying from DC to 1 MHZ, high frequencies favoring the formation of particle chains connecting the electrodes. Currents were monitored with a potentiostat and displayed on an oscilloscope. For convenience, epi-fluorescence as well as reflection differential interference contrast microscopy employed laser illumination. Light-induced modulations in EIS impedance were also produced with a simple 100W microscope illuminator as well as with a 3mW laser diode emitting light at 650-680 nm.

30 Colloidal beads, both anionic and cationic as well as nominally neutral, with a diameter in the range from several hundred Angstroms to 20 microns, stored in a NaN<sub>2</sub> solution, were employed.

5 Close attention was paid to colloidal stability to avoid non-specific interactions between particles and between particles and the electrode surface. Bacterial contamination of colloidal suspensions was scrupulously avoided.

10 Typical operating conditions producing, unless otherwise indicated, most of the results described herein, were: 0.2 mM NaN<sub>3</sub> (sodium azide) solutions, containing particles at a concentration so as to produce not more than a complete monolayer of particles when deposited on the electrode; applied DC potentials in the range of 1-4V, and AC potentials in the range of 1-10V (peak-to-peak) and 500Hz - 10kHz, with an electrode gap of 50 microns; anionic (carboxylated polystyrene) beads of 2 micron diameter, as well as (nominally neutral) polystyrene beads of 2-20 micron diameter.

15 The method and apparatus of the present invention may be used in several different areas, examples of which are discussed in detail. Each example includes background information followed by the application of the present invention to that particular application.

20

#### 25 Example I - Fabrication of Surfaces and Coatings with Designed Properties

30 The present invention may be used to fabricate planar surfaces and coatings with designed properties. Specifically, the functional elements of the present invention enable the formation of arrays composed of particles of a wide range of sizes (approximately 100 Angstrom to 10 microns) and chemical composition or surface functionality in response to AC or DC electric fields. These arrays may be placed and delineated in designated areas of the substrate, and the interparticle spacing and internal state of order within the array may be controlled by adjusting the applied field prior to anchoring the array to the substrate. The newly formed surfaces display pre-designed mechanical, optical and chemical characteristics, and they may be subjected to further modification by subsequent treatment such as chemical cross-linking.

35 The mechanical and/or chemical modification of surfaces and coatings principally determines the interaction between materials in a wide range of applications that depend on low adhesion (e.g., the familiar "non-stick" surfaces important in housewares) or low friction (e.g., to reduce wear in computer hard disks), hydrophobicity (the tendency to repel water, e.g., of certain fabrics), catalytic activity or specific chemical functionality to either suppress molecular interactions with surfaces or

55

5 to promote them. The latter area is of particular importance to the development of  
reliable and durable biosensors and bioelectronic devices. Finally, a large number of  
10 applications depend on surfaces of defined topography and/or chemical functionality to  
act as templates controlling the growth morphology of deposited materials or as  
5 "command surfaces" directing the alignment of optically active molecules in deposited  
thin organic films, as in liquid crystal display applications.

15 Extensive research has been devoted to the formation of surfaces by  
adsorption of thin organic films of known composition from the liquid or gas phase by  
several methods. Notwithstanding their seeming simplicity and wide-spread use, these  
10 methods can be difficult to handle in producing reliable and reproducible results. In  
20 addition, molecular films are not well suited to produce surfaces displaying a regular  
topography.

25 An alternative approach to the problem is the modification of conductive  
surfaces by electrophoretic deposition of suspended particulates. This is a widely used  
15 technique in industrial settings to produce paint coatings of metal parts, and to deposit  
phosphor for display screens. The active deposition process significantly enhances the  
kinetics of formation (in contrast to passive adsorption of organic films from solution),  
30 an important consideration in practical applications. Electrophoretic deposition requires  
high DC electric fields and produces layers in which particles are permanently adsorbed  
20 to the surface. While particles in so-deposited monolayers are usually randomly  
distributed, the formation of polycrystalline monolayers of small (150 Angstrom) gold  
35 colloids on carbon-coated copper grids is also known. However, the use of carbon-  
coated copper grids as substrates is not desirable in most applications.

40 Prior art methods have been described for the formation of ordered planar  
25 arrays of particles under certain conditions. For example, the formation of ordered  
colloidal arrays in response to AC electric fields on conductive indium tin oxide (ITO)  
electrodes is known. However, the resulting layers were composed of small patches of  
45 ordered arrays, randomly distributed over the surface of the otherwise bare ITO  
substrate. Arrays of monodisperse colloidal beads and globular proteins also have been  
30 previously fabricated by using convective flow and capillary forces. However, this  
latter process has the disadvantage of leaving deposited particle arrays immobilized and  
50 exposed to air, making it difficult to modify arrays by subsequent liquid phase

chemistry.

The present invention provides a method of forming planar arrays with precise control over the mechanical, optical and chemical properties of the newly created layer. This method has several distinct advantages over the prior art. These result from the combination of AC electric field-induced array formation on insulating electrodes (Si/SiO<sub>x</sub>) that are patterned by UV-mediated oxide regrowth. The process of the present invention enables the formation of ordered planar arrays from the liquid phase (in which particles are originally suspended) in designated positions, and in accordance with a given overall outline. This eliminates the above-stated disadvantages of the prior art, i.e., dry state, irregular or no topography, random placement within an aggregate, immobilization of particles and uncontrolled, random placement of ordered patches on the substrate.

An advantage of the present invention is that arrays are maintained by the applied electric field in a liquid environment. The process leaves the array in a state that may be readily disassembled, subjected to further chemical modification such as cross-linking, or made permanent by chemical anchoring to the substrate. Furthermore, the liquid environment is favorable to ensure the proper functioning of many proteins and protein supramolecular assemblies of which arrays may be composed. It also facilitates the subsequent liquid-phase deposition of additional layers of molecules (by chemical binding to beads or proteins in the deposited layer), the cycling of arrays between states of different density and internal order (including complete disassembly of the array) in response to electric fields and the chemical cross-linking of particles into two-dimensionally connected layers, or gels, formed, for example, of chemically functionalized silica spheres. The present invention can be practiced on insulating electrodes such as oxide-capped silicon, to minimize Faradaic processes that might adversely affect chemical reactions involved in the gelation process or in anchoring the array to the substrate. The use of Si/SiO<sub>x</sub> electrodes also enables the control of array placement by external illumination.

The formation of colloidal arrays composed of small particles in accordance with the present invention provides a route to the fabrication of surfaces with relief structure on the scale of the particle diameter. Aside from their optical properties, such "micro-rough" surfaces are of interest as substrates for the deposition of DNA in

5 such a way as to alleviate steric constraints and thus to facilitate enzyme access.

10 Particles to which the invention applies include silica spheres, polymer colloids, lipid vesicles (and related assemblies) containing membrane proteins such as bacteriorhodopsin (bR) a light-driven proton pump that can be extracted in the form of  
5 membrane patches and disks or vesicles. Structured and functionalized surfaces composed of photoactive pigments are of interest in the context of providing elements of planar optical devices for the development of innovative display and memory  
15 technology. Other areas of potential impact of topographically structured and chemically functionalized surfaces are the fabrication of template surfaces for the controlled nucleation of deposited layer growth and command surfaces for liquid crystal  
20 alignment. The present invention also enables the fabrication of randomly heterogeneous composite surfaces. For example, the formation of arrays composed of a mixture of hydrophobic and hydrophilic beads of the same size creates a surface whose wetting and lubrication characteristics may be controlled by the composition of the  
25 deposited mixed bead array. In this way, the location of the individual beads is random, but the relative proportion of each type of bead within the array is controllable.

#### 30 Example II - Assembly of Lens Arrays and Optical Diffraction Elements

The present invention can be used to fabricate lens arrays and other  
20 surface-mounted optical elements such as diffraction gratings. The functional elements of the present invention enable the placement and delineation of these elements on ITO, facilitating integration with existing planar display technology, and on Si/SiO<sub>x</sub>,  
35 facilitating integration with existing silicon-based device technology.

40 Silica or other oxide particles, polymer latex beads or other objects of high refractive index suspended in an aqueous solution, will refract light. Ordered  
25 planar arrays of beads also diffract visible light, generating a characteristic diffraction pattern of sharp spots. This effect forms the basis of holographic techniques in optical  
45 information processing applications.

30 A. - The present invention provides for the use of arrays of refractive colloidal beads as light collection elements in planar array formats in conjunction with  
50 low light level detection and CCD imaging. CCD and related area detection schemes



5 will benefit from the enhanced light collection efficiency in solid-phase fluorescence or luminescence binding assays.

10 This assay format relies on the detection of a fluorescence signal indicating the binding of probes to bead-anchored targets in the vicinity of the detector.  
5 To maximize through-put, it is desirable to monitor simultaneously as many binding events as possible. It is here that array formation by the methods of the present invention is particularly valuable because it facilitates the placement and tight packing of  
15 beads in the target area monitored by the CCD detector, while simultaneously providing for the additional benefit of lensing action and the resulting increase in light collection  
20 efficiency.

25 Increased collection efficiency has been demonstrated in experiments employing individual, large (10 micron diameter) polystyrene beads as lensing elements to image small (1 micron diameter) fluorescent polystyrene beads. Under the experimental conditions set forth above an applied voltage of 5V (pp) at 300 Hz induced  
30 the collection of small particles under individual large beads within a second. This is shown in Fig. 5, where small beads alone, e.g., 52, appear dim, whereas small beads, e.g., 54, gathered under a large bead 56 appear brighter and magnified. The small beads redisperse when the voltage is turned off.

35 B. - The use of colloidal bead arrays as diffraction gratings and thus as holographic elements is known. Diffraction gratings have the property of diffracting light over a narrow range of wavelengths so that, for given angle of incidence and wavelength of the illuminating light, the array will pass only a specific wavelength (or a narrow band of wavelengths centered on the nominal value) that is determined by the  
40 inter-particle spacing. Widely discussed applications of diffraction gratings range from  
25 simple wavelength filtering to the more demanding realization of spatial filters and related holographic elements that are essential in optical information processing.

45 The present invention provides for a rapid and well controlled process of forming planar arrays in a state of crystalline order which will function as surface-mounted optical diffraction elements. In addition, the resulting surfaces may be  
30 designed to display topographical relief to enhance wave-length selective reflectivity. These arrays may be formed in designated areas on a substrate surface. In contrast to  
50 the slow and cumbersome prior art method of fabricating such arrays by way of forming

5 equilibrium crystals in aqueous solutions of low salt content, the present invention  
provides a novel approach to rapidly and reliably fabricate particle arrays at a  
solid-liquid interface. This approach relies on field-induced formation of arrays to  
10 trigger the process, and on UV-mediated patterning or light control to position and shape  
the arrays. In addition, the inter-particle distance, and internal state of order, and hence  
5 the diffraction characteristics of the array, may be fine-tuned by adjusting the applied  
electric field. For example, a field-induced, reversible order-disorder transition in the  
15 array will alter the diffraction pattern from one composed of sharp spots to one  
composed of a diffuse ring. The assembly of such arrays on the surface of silicon  
20 wafers, as described herein, provides a direct method of integration into existing  
microelectronic designs. Arrays may be locked in place by chemical coupling to the  
substrate surface, or by relying on van der Waals attraction between beads and substrate.

#### 25 Example III - A Novel Mechanism for the Realization of a Particle-Based Display

15 The present invention provides the elements to implement lateral particle  
motion as a novel approach to the realization of a particle-based display. The elements  
of the present invention provide for the control of the lateral motion of small particles in  
30 the presence of a pre-formed lens array composed of large, refractive particles.

Colloidal particulates have been previously employed in flat-panel display  
20 technology. The operating principle of these designs is based on electrophoretic motion  
of pigments in a colored fluid confined between two planar electrodes. In the OFF  
35 (dark) state, pigments are suspended in the fluid, and the color of the fluid defines the  
appearance of the display in that state. To attain the ON (bright) state, particles are  
assembled near the front (transparent) electrode under the action of an electric field. In  
40 this latter state, incident light is reflected by the layer of particles assembled near the  
electrode, and the display appears bright. Prototype displays employing small reflective  
25 particles in accordance with this design are known. However, these displays suffered  
from a number of serious problems including: electrochemical degradation and lack of  
45 colloidal stability as a result of prolonged exposure to the high DC electric fields  
required to achieve acceptable switching speeds; and non-uniformities introduced by  
30 particle migration in response to field gradients inherent in the design of the addressing  
scheme.

5 The present invention provides a novel mechanism for the design of a  
particle-based display which takes advantage of electric field-induced array formation as  
well as controlled, field-induced lateral particle displacements. First, a lens array  
10 composed of colloidal beads is formed. This lens array also serves as a spacer array to  
5 maintain a well-defined gap between the bottom electrode and the top electrode that may  
now be placed over the (pre-formed) array. This facilitates fabrication of uniform flat  
panel displays with a narrow gap that is determined by the particle diameter.

15 Next, small colloidal particles are added to the electrolyte solution in the  
gap. These may be fluorescent, or may be reflecting incident white light. Under the  
20 action of an AC electric field of appropriate frequency, these small particles can be  
moved laterally to assemble preferentially within the footprint of a larger bead. When  
viewed through a larger bead, small fluorescent beads assembled under a large bead  
appear bright as a result of the increased light collection efficiency provided by the  
25 lensing action of the large bead; this is the ON state (Fig. 5). When moved outside the  
15 footprint of the larger bead, particles appear dim, and may be made entirely invisible by  
appropriate masking; this is the OFF state. The requisite lateral particle motion may be  
induced by a change in the applied voltage or a change in light intensity. Each large or  
30 lensing bead introduces a lateral nonuniformity in the current distribution within the  
electrolyte because the current is perturbed by the presence of each lensing bead.

20 In contrast to the prior art displays, the present invention employs AC,  
35 not DC fields, and insulating (rather than conductive) electrodes, thereby minimizing  
electrochemical degradation. The lateral non-uniformity introduced by the lens array is  
desirable because it introduces lateral gradients in the current distribution within the  
display cell. These gradients mediate the lateral motion of small beads over short  
40 characteristic distances set by the diameter of the large lensing beads, to effect a  
25 switching between ON and OFF states. Thus, the present invention readily  
accommodates existing technology for active matrix addressing.

#### 45 Example IV - Separation and Sorting of Beads and Particles

30 The present invention can be used to implement several procedures for the  
separation and sorting of colloidal particles and biomolecules in a planar geometry.  
50 Specifically, these include techniques of lateral separation of beads in mixtures.

Individual beads may be removed from an array formed in response to an electric field by the application of optical tweezers.

The separation of components in a given mixture of chemical compounds is a fundamental task of analytical chemistry. Similarly, biochemical analysis frequently calls for the separation of biomolecules, beads or cells according to size and/or surface charge by electrophoretic techniques, while the sorting (most commonly into just two sub-classes) of suspended cells or whole chromosomes according to optical properties such as fluorescence emission is usually performed using field-flow fractionation including flow cytometry and fluorescence-activated cell sorting.

In a planar geometry, bead mixtures undergoing diffusion have been previously separated according to mobility by application of an AC electric field in conjunction with lithographic patterning of the electrode surface designed to promote directional drift. Essentially, as described in U. S. Patent No. 5,593,565 to Ajdari et al., the contents of which are included herein by reference, the AC or pulsing electric field is used to move small beads in a particular direction over a period of time, advancing beads of higher mobility relative to those of lower mobility. Capillary electrophoresis has been implemented in a planar geometry, see e.g., B.B. Haab and R.A. Mathies, Anal. Chem 67, 3253-3260 (1995), the contents of which are incorporated herein by reference.

The methods of the present invention may be applied in several ways to implement the task of separation, sorting or isolation in a planar geometry. In contrast to the prior art approaches, the present invention provides a significant degree of flexibility in selecting from among several available procedures, the one best suited to the particular task at hand. In some cases, more than one separation technique may be applied, and this provides the basis for the implementation of two-dimensional separation. That is, beads may be separated according to two different physical-chemical characteristics. For example, beads may first be separated by size and subsequently, by raising the applied frequency to induce chain formation, by polarizability. This flexibility offers particular advantages in the context of integrating analytical functionalities in a planar geometry. Several techniques will now be described.

- i) The present invention may be used to implement "sieving" in

5 lateral, electric field-induced flow on surfaces patterned by UV-mediated oxide regrowth  
to sort beads in a mixture by size. The fundamental operations of the invention are  
invoked to set up directed lateral particle motion along conduits laid out by UV-mediated  
10 oxide regrowth. Conduits are designed to contain successively narrower constrictions  
5 through which particles must pass. Successively finer stages allow only successively  
smaller particles to pass in this "sieving" mechanism (Fig. 9a). As shown in Fig. 9a,  
15 the primary particle flow is in the direction left to right, while a transverse flow is  
established in the top to bottom direction utilizing an oxide profile as shown.  
Additionally, rows of barriers 92 made from thick oxide are positioned along the conduit  
20 with the spacing between the barriers in each row decreasing in the transverse direction.  
As the particles move along the conduit, the rows of barriers act to separate out smaller  
particles in the transverse direction. In contrast to previous methods based on  
electrophoretic separation, large DC electric fields, and the attendant potential problem  
25 of electrolysis and interference from electroosmotic flow in a direction opposite to the  
15 field-directed particle transport, the present invention uses AC electric fields and lateral  
gradients in interfacial impedance to produce transport. The present method has the  
advantage of avoiding electrolysis and it takes explicit advantage of electroosmotic flow  
30 to produce and control particle transport.

In addition, the use of Si/SiO<sub>x</sub> electrodes enables the use of the light-  
20 control component of the present invention to modify lateral transport of beads in real  
time. For example, external illumination may be employed to locally neutralize the  
35 lateral impedance gradient induced by UV-mediated oxide regrowth. Particles in these  
neutral "zones" would no longer experience any net force and come to rest. This  
principle may be used as a basis for the implementation of a scheme to locally  
40 concentrate particles into sharp bands and thereby to improve resolution in subsequent  
25 separation.

ii) The present invention may be used to implement "zone refining",  
45 a process of excluding minority components of a mixture by size or shape from a  
growing crystalline array of majority component. This process explicitly depends on the  
30 capabilities of the present invention to induce directional crystallization.

The process of zone refining is employed with great success in producing  
50 large single crystals of silicon of very high purity by excluding impurities from the host

5 lattice. The concept is familiar from the standard chemical procedure of purification by  
re-crystallization in which atoms or molecules that are sufficiently different in size,  
10 shape or charge from the host species so as not to fit into the forming host crystal lattice  
as a substitutional impurity, are ejected into solution.

5 By enabling the growth of planar arrays, in a given direction and at a  
controlled rate, the present invention facilitates the implementation of an analogous zone  
refining process for planar arrays. The most basic geometry is the linear geometry. A  
15 multi-component mixture of beads of different sizes and/or shapes is first captured in a  
rectangular holding area on the surface, laid out by UV-patterning. Next, crystallization  
20 is initiated at one end of the holding area by illumination and allowed to slowly advance  
across the entire holding area in response to an advancing pattern of illumination. In  
general, differences of approximately 10% in bead radius trigger ejection.

25 iii) The present invention may be used to implement fractionation in a  
transverse flow in a manner that separates particles according to mobility.

15 Field-flow fractionation refers to an entire class of techniques that are in  
wide use for the separation of molecules or suspended particles. The principle is to  
separate particles subjected to fluid flow in a field acting transverse to the flow. A  
30 category of such techniques is subsumed under the heading of electric-field flow  
fractionation of which free-flow electrophoresis is a pertinent example because it is  
20 compatible with a planar geometry. Free-flow electrophoresis employs the continuous  
flow of a replenished buffer between two narrowly spaced plates in the presence of a DC  
35 electric field that is applied in the plane of the bounding plates transverse to the direction  
of fluid flow. As they traverse the electric field, charged particles are deflected in  
proportion to their electrophoretic mobility and collected in separate outlets for  
40 subsequent analysis. In contrast to conventional electrophoresis, free-flow  
25 electrophoresis is a continuous process with high throughput and it requires no  
supporting medium such as a gel.

45 The present invention enables the implementation of field-flow  
fractionation in a planar geometry. As previously discussed herein, impedance gradients  
30 imposed by UV-oxide profiling serve to mediate particle motion along the electrode  
surface in response to the external electric field. In a cell with a narrow gap, the  
50 resulting electrokinetic flow has a "plug" profile and this has the advantage of exposing

all particles to identical values of the flow velocity field, thereby minimizing band distortions introduced by the parabolic velocity profile of the laminar flow typically employed in free-flow electrophoresis.

A second flow field, transverse to the primary flow direction, may be employed to mediate particle separation. This deflecting flow may be generated in response to a second impedance gradient. A convenient method of imposing this second gradient is to take advantage of UV-oxide patterning to design appropriate flow fields. Both longitudinal and transverse flow would be recirculating and thus permit continuous operation even in a closed cell, in contrast to any related prior art technique.

Additional flexibility is afforded by invoking the light-control component of the present invention to illuminate the substrate with a stationary pattern whose intensity profile in the direction transverse to the primary fluid flow is designed to induce the desired impedance gradient and hence produce a transverse fluid flow. (Fig. 9b). This has the significant advantage of permitting selective activation of the transverse flow in response to the detection of a fluorescent bead crossing a monitoring window upstream. Non-fluorescent beads would not activate the transverse flow and would not be deflected. This procedure represents a planar analog of flow cytometry, or fluorescence-activated cell sorting.

iv) The invention may be used to induce the formation of particle chains in the direction normal to the plane of the electrode. The chains represent conduits for current transport between the electrodes and their formation may reflect a field-induced polarization. Chains are much less mobile in transverse flow than are individual particles so that this effect may be used to separate particles according to the surface properties that contribute to the net polarization. The effect of reversible chain formation has been demonstrated under the experimental conditions stated herein. For example, the reversible formation of chains occurs, for carboxylated polystyrene beads of 1 micron diameter, at a voltage of 15 V (pp) at frequencies in excess of 1MHz.

v) The invention may be used to isolate individual beads from a planar array.

Fluorescence binding assays in a planar array format, as described herein, may produce singular, bright beads within a large array, indicating particularly strong binding. To isolate and retrieve the corresponding beads, optical tweezers in the form

5 of a sharply focused laser spot, may be employed to lock onto an individual bead of interest. The light-control component of the present invention may be used in conjunction with the optical tweezers to retrieve such an individual bead by moving the  
10 array relative to the bead, or vice versa, or by disassembling the array and retaining only the marked bead. This is a rather unique capability that will be particularly useful in the context of isolating beads in certain binding assays.

15 Commercial instrumentation is available to position optical tweezers in the field of a microscope. Larger scale motion is facilitated by translocating the array in-situ or simply by moving the external sample fixture. This process lends itself to  
20 automation in conjunction with the use of peak-finding image analysis software and feedback control.

vi) The invention may be used to implement a light-induced array sectioning ("shearing") operation to separate fluorescent, or otherwise delineated  
25 portions of an array from the remainder. This operation makes it possible to segment a given array and to isolate the corresponding beads for downstream analysis.

The basis for the implementation of this array segmentation is the light-control component of the present invention, in the mode of driving particles from  
30 an area of a Si/SiO<sub>x</sub> interface that is illuminated with high intensity. It is emphasized here that this effect is completely unrelated to the light-induced force on beads that underlies the action of optical tweezers. The present effect which operates on large sets  
35 of particles, was demonstrated under the experimental conditions stated herein using a 100W illuminator on a Zeiss UEM microscope operated in epi-illumination. A simple implementation is to superimpose, on the uniform illumination pattern applied to the entire array, a line-focussed beam that is positioned by manipulation of beam steering  
40 elements external to the microscope. Beads are driven out of the illuminated linear portion. Other implementations take advantage of two separately controlled beams that are partially superimposed. The linear sectioning can be repeated in different relative  
45 orientations of shear and array.

#### 30 Example V - Fabrication of Spatially Encoded Bead Arrays

50 The present invention provides a method to transfer suspensions of beads or biomolecules to the electrode surface in such a way as to preserve the spatial



5 encoding in the original arrangement of reservoirs, most commonly the conventional  
8x12 arrangement of wells in a microtiter plate. Such a fluid transfer scheme is of  
significant practical importance given that compound libraries are commonly handled  
10 and shipped in 8x12 (or equivalent) wells.

5 The present invention utilizes chemical patterning to define individual  
compartments for each of MxN sets of beads and confine them accordingly. In the  
present instance, patterning is achieved by UV-mediated photochemical oxidation of a  
15 monolayer of thiol-terminated alkylsilane that is chemisorbed to the Si/SiO<sub>x</sub> substrate.  
Partial oxidation of thiol moieties produces sulfonate moieties and renders the exposed  
20 surface charged and hydrophilic. The hydrophilic portions of the surface, in the form of  
a grid of squares or circles, will serve as holding areas.

In accordance with the present invention, the first function of  
surface-chemical patterning into hydrophilic sections surrounded by hydrophobic  
25 portions is to ensure that droplets, dispensed from different wells, will not fuse once  
they are in contact with the substrate. Consequently, respective bead suspensions will  
remain spatially isolated and preserve the lay-out of the original MxN well plate. The  
second role of the surface chemical patterning of the present invention is to impose a  
30 surface charge distribution, in the form of the MxN grid pattern, which ensures that  
individual bead arrays will remain confined to their respective holding areas even as the  
20 liquid phase becomes contiguous.

35 The layout-preserving transfer procedure involves the steps illustrated in  
Figs. 6a-c. First, as shown in side view in Fig. 6a, the MxN plate of wells 62 is  
registered with the pattern 64 on the planar substrate surface. Well bottoms 62, are  
pierced to allow for the formation of pendant drops of suspension or, preferably, the  
40 process is facilitated by a fixture (not shown) providing MxN effective funnels to match  
the geometric dimensions of the MxN plate on the top and reduce the size of the  
dispensing end. Such a dispensing fixture will also ensure the precise control of droplet  
45 volumes, adjusted so as to slightly overfill the target holding area on the patterned  
substrate surface. The set of MxN drops is then deposited by bringing them in contact  
30 with the hydrophilic holding areas of the pre-patterned substrate and relying on capillary  
action.

50 Next, the plate is retracted, and the top electrode is carefully lowered to

5 form the electrochemical cell, first making contact as shown in Fig. 6b, with individual liquid-filled holding areas on the substrate to which suspensions are confined.

10 Overfilling ensures that contact is made with individual suspensions. The electric field is now turned on to induce array formation in the MxN holding areas and to ensure the  
5 preservation of the overall configuration of the MxN sets of beads while the gap is closed further (or filled with additional buffer) to eventually fuse individual droplets of suspension into a contiguous liquid phase as shown in Fig. 6c. In the fully assembled  
15 cell of Fig. 6c, while the droplets are fused together, the beads from each droplet are maintained in and isolated in their respective positions, reflecting the original MxN  
20 arrangement of wells. The present invention thus provides for the operations required in this implementation of a layout-preserving transfer procedure to load planar electrochemical cells.

#### 25 Example VI - Fabrication of Dynamic Planar Bead Arrays for Parallel Assays

15 The present invention provides a method to produce a heterogeneous panel of beads and potentially of biomolecules for presentation to analytes in an adjacent liquid. A heterogeneous panel contains particles or biomolecules which differ in the  
30 nature of the chemical or biochemical binding sites they offer to analytes in solution. The present method relies on the functional elements of the invention to assemble a  
20 planar array of a multi-component mixture of beads which carry chemical labels in the form of tag molecules and may be so identified subsequent to performing the assay. In  
35 the event of binding, the analyte is identified by examination of the bead, or cluster of beads, scoring positive.

40 Diagnostic assays are frequently implemented in a planar format of a  
25 heterogeneous panel, composed of simple ligands, proteins and other biomolecular targets. For example, in a diagnostic test kit, a heterogeneous panel facilitates the rapid testing of a given analyte, added in solution, against an entire set of targets. Heterogeneous panels of proteins are of great current interest in connection with the emerging  
45 field of proteome research. The objective of this research is to identify, by scanning the  
30 panel with sensitive analytical techniques such as mass spectrometry, each protein in a multi-component mixture extracted from a cell and separated by two-dimensional gel  
50 electrophoresis. Ideally, the location of each spot uniquely corresponds to one particular

5 protein. This analysis would permit, for example, the direct monitoring of gene expression levels in a cell during a particular point in its cycle or at a given stage during embryonic development.

10 The fabrication of an array of heterogeneous targets is central to recently proposed strategies of drug screening and DNA mutation analysis in a planar format. The placement of ligands in a specific configuration on the surface of a planar substrate serves to maintain a key to the identity of any one in a large set of targets presented simultaneously to an analyte in solution for binding or hybridization. In an assay relying on fluorescence, binding to a specific target will create bright spots on the substrate  
15 whose spatial coordinates directly indicate the identity of the target.

20 Three principal strategies have been previously employed to fabricate heterogeneous panels. First, protein panels may be created by two-dimensional gel electrophoresis, relying on a DC electric field to separate proteins first by charge and then by size (or molecular weight). Even after many years of refinement, this technique  
25 yields results of poor reproducibility which are generally attributed to the poorly defined properties of the gel matrix.

30 Second, individual droplets, drawn from a set of reservoirs containing solutions of the different targets, may be dispensed either by hand or by employing one of several methods of automated dispensing (or "printing"; see e.g., Schena et al.,  
35 Science 270, 467-470 (1995), the contents of which are incorporated herein by reference). Printing has been applied to create panels of oligonucleotides intended for screening assays based on hybridization. Printing leaves a dried sample and may thus not be suitable for proteins that would denature under such conditions. In addition, the attendant fluid handling problems inherent in maintaining, and drawing samples from a  
40 large number of reservoirs are formidable.

45 Third, target ligands may be created by invoking a variant of solid phase synthesis based on a combinatorial strategy of photochemically activated elongation reactions. This approach has been limited by very formidable technical problems in the chemical synthesis of even the simplest, linear oligomers. The synthesis of non-linear  
50 compounds in this planar geometry is extremely difficult.

55 The present invention of forming heterogeneous panels requires the chemical attachment of target ligands to beads. Ligands may be coupled to beads

5 “off-line” by a variety of well established coupling reactions. For present purposes, the  
bead identity must be chemically encoded so it may be determined as needed. Several  
10 methods of encoding, including binary encoding, of beads are available. For example,  
short oligonucleotides may serve the purpose of identifying a bead via their sequence  
5 which may be determined by microscale sequencing techniques. Alternatively,  
chemically inert molecular tags may be employed that are readily identified by standard  
15 analytical techniques.

In contrast to all prior art methods, the present invention provides a novel  
method to create heterogeneous panels by in-situ, reversible formation of a planar array  
20 of chemically encoded beads in solution adjacent to an electrode. The array may be  
random with respect to chemical identity but is spatially ordered. This procedure offers  
several advantages. First, it is reversible so that the panel may be disassembled  
following the binding assay to discard beads scoring negative. Positive beads may be  
25 subjected to additional analysis without the need for intermediate steps of sample  
retrieval, purification or transfer between containers. Second, the panel is formed when  
needed, that is, either prior to performing the actual binding assay, or subsequent to  
performing the assay on the surface of individual beads in suspension. The latter mode  
30 minimizes potential adverse effects that can arise when probes bind to planar target  
surfaces with a high concentration of target sites. Third, to accommodate optical  
20 analysis of individual beads, interparticle distances within the array may be adjusted by  
field-induced polarization or by the addition of inert spacer particles that differ in size  
35 from the encoded beads. Fig. 7 shows the use of small spacer beads 72 for separating  
encoded beads 74. As shown, the spacing of beads 74 is greater than the spacing of  
comparable beads in Fig. 4b. Finally, UV-mediated oxide regrowth, as provided by the  
40 present invention, readily facilitates the embedding of a grid pattern of selected  
25 dimension into the substrate to ensure the formation of small, layout-preserving  
subarrays in the low-impedance fields of the grid.

45 To create the panel, a multi-component mixture of beads carrying, for  
example, compounds produced by bead-based combinatorial chemistry, is placed  
30 between electrodes. Each type of bead may be present in multiple copies. Arrays are  
formed in response to an external field in a designated area of the electrode surface.  
50 This novel approach of in-situ assembly of panels relies on beads that carry a unique

5 chemical label, or code, to permit their identification subsequent to the completion of a  
binding assay. Alternatively, beads may be marked ("painted") on-line by way of a  
photochemical bead-coloring method. Selected beads in an array are individually  
10 illuminated by a focused light source to trigger a coloring reaction on the bead surface or  
in the bead interior to indicate a positive assay score. Beads so marked can be  
subsequently separated from unmarked beads by a light-activated sorting method  
described herein. Numerous UV-activated reactions are available to implement this  
15 bead-coloring method.

The present invention provides for several methods of discarding beads  
20 with negative scores, typically the vast majority, while retaining those with positive  
scores. This method take advantage of the fact that, in contrast to all prior art methods,  
the array represents a temporary configuration of particles that is maintained by the  
applied electric field and may be rearranged or disassembled at will. This capability,  
25 along with the fact that biomolecules are never exposed to air (as in the prior art method  
of printing) facilitates the in-situ concatenation of analytical procedures that require the  
heterogeneous panel in conjunction with subsequent, "downstream" analysis.

First, if positive beads are clustered in a subsection of the array, the  
30 light-controlled array splitting operation of the present invention may be invoked to  
dissect the array so as to discard negative portions of the array (or recycle them for  
subsequent use). Second, if positive and negative beads are randomly interspersed, a  
35 fluorescence-activated sorting method, implemented on the basis of the present invention  
in a planar format, as described herein, may be invoked. In the case of fluorescence-  
activated sorting, positive and negative beads may be identified as bright and dark  
objects, respectively. In the special case that only a few positive beads stand out, these  
40 may be removed from the array by locking onto them with optical tweezers, a tool to  
trap and/or manipulate individual refractive particles under illumination, and  
disassembling the array by removing the field, or subjecting the entire array to lateral  
displacement by the fundamental operations of the present invention.  
45

The typical task in screening a large set of compounds is one of looking  
30 for a very small number of positive events in a vast number of tests. The set of  
discarded beads will typically involve the majority at each stage in the assay. The  
50 procedure of the present invention therefore minimizes the effort invested in negative

5 events, such as the challenging in-situ synthesis of target ligands irrespective of whether or not they will prove to be of interest by binding a probe offered in solution.

10 The method of forming a heterogeneous panel according to the present invention contains beads of each type in generally random assembly. The creation of a  
5 heterogeneous panel with each position in the panel containing a cluster of beads of the same type, that is, beads originating in the same reservoir (Fig. 6a), may be desirable so as to ensure a sufficiently large number of positive events to facilitate detection. A  
15 practical solution follows from the application of the layout-preserving fluidic transfer scheme described herein. In this procedure, beads from an MxN well plate are  
10 transferred layout-preservingly onto a chemically patterned substrate in such a way as to preserve the spatial encoding of bead identities.

#### Example VII - Binding and Functional Assays in Planar Bead Array Format

25 The present invention can be used to implement mixed-phase binding assays as well as certain functional assays in a planar array format. Several  
15 combinations are possible reflecting the presence of probe or target in solution, on the surface of colloidal beads, or on the electrode surface. The methods of the present  
30 invention facilitate the formation of a planar array to present targets to probes in solution prior to performing the binding assay ("pre-formed" array; Fig. 8). Alternatively, a  
20 planar array of beads may be formed in front of a detector surface subsequent to performing the binding assay in suspension ("post-formed" array; Fig. 8). The present  
35 invention also provides the methods to implement functional assays by enabling the assembly of certain cell types adjacent to a planar detector or sensor surface to monitor the effects of exposure of the cells to small molecule drugs in solution.

40 Binding assays, particularly those involving proteins such as enzymes and antibodies, represent a principal tool of medical diagnostics. They are based on the  
45 specific biochemical interaction between a probe, such as a small molecule, and a target, such as a protein. Assays facilitate the rapid detection of small quantities of an analyte in solution with high molecular specificity. Many procedures have been designed to  
30 produce signals to indicate binding, either yielding a qualitative answer (binding or no binding) or quantitative results in the form of binding or association constants. For  
50 example, when an enzyme binds an analyte, the resulting catalytic reaction may be used

5 to generate a simple color change to indicate binding, or it may be coupled to other  
processes to produce chemical or electrical signals from which binding constants are  
determined. Monoclonal antibodies, raised from a single common precursor, may be  
10 prepared to recognize virtually any given target, and immunoassays, based on  
5 antibody-antigen recognition and binding, have developed into an important diagnostic  
tool. As with enzyme binding, antibody binding of an antigenic analyte may be detected  
15 by a variety of techniques including the classic method of enzyme-linked immunoassays  
(ELISA) in which the reaction of an antibody-coupled enzyme is exploited as an  
indicator. A common and conceptually simple scheme ensures the detection of antibody  
20 binding to a target analyte by supplying a fluorescently labeled second antibody that  
recognizes the first (or primary) antibody.

Binding assays involving soluble globular proteins are often performed in  
solution to ensure unbiased interactions between protein and target. Such liquid phase  
25 assays, especially when performed at low concentrations of target or probe, minimize  
15 potential difficulties that may arise when either target or probe are present in abundance  
or in close proximity. By the same token, the kinetics tend to be slow. Cooperative  
effects, such as crowding, arising from the close proximity of probes must be carefully  
30 controlled when either probe or target is chemically anchored to a solid substrate.

Nonetheless, this latter solid phase format of binding assays is also very  
20 commonly employed whenever the situation demands it. For example, the presence of a  
protein on the surface of a cell may be exploited in "panning" for the cells that express  
35 this protein in the presence of many other cells in a culture that do not: desired cells  
attach themselves to the surface of a container that is pre-coated with a layer of a  
secondary antibody directed against a primary antibody decorating the desired  
40 cell-surface protein. Similarly, certain phages may be genetically manipulated to display  
25 proteins on their surface, and these may be identified by a binding assay involving a  
small molecule probe such as an antigen if the protein displayed is an antibody (Watson  
45 et al., "Recombinant DNA", 2nd Edition (Scientific American Books, W.H. Freeman  
and Co., New York, NY, 1983), the contents of which are incorporated herein by  
30 reference). In addition, the planar geometry accommodates a variety of optical and  
electrical detection schemes implemented in transducers and sensors.

A combination of liquid phase and solid phase assay may be developed by

5 using beads that are decorated with either probe or target, as in procedures that employ decorated magnetic beads for sample preparation or purification by isolating binding  
10 from non-binding molecules in a given multi-component mixture. Recent examples of the use of these beads include the purification of templates for DNA sequencing  
15 applications or the extraction of mRNAs from (lysed) cells by hybridization to beads that are decorated with poly-adenine (polyA) residues.

15 Functional assays involving suitable types of cells are employed to monitor extracellular effects of small molecule drugs on cell metabolism. Cells are placed in the immediate vicinity of a planar sensor to maximize the local concentration  
20 of agents released by the cell or to monitor the local pH.

20 The present invention provides the means to implement mixed phase binding assays in a planar geometry with a degree of flexibility and control that is not available by prior art methods. Thus, it offers the flexibility of forming, in-situ,  
25 reversibly and under external spatial control, either a planar panel of target sites for binding of analyte present in an adjacent liquid phase, or a planar array of probe-target  
15 complexes subsequent to performing a binding assay in solution. Binding may take place at the surface of individual beads suspended in solution, at the surface of beads  
30 pre-assembled into arrays adjacent to the electrode surface, or at the electrode surface itself. Either the target or probe molecule must be located on a bead to allow for a bead-  
20 based assay according to the present invention. As shown in Fig. 8, if the probe molecule P is located on a bead, then the target molecule T may be either in solution, on  
35 a bead or on the electrode surface. The converse is also true.

40 For example, the methods of the present invention may be used to implement panning, practiced to clone cell surface receptors, in a far more expeditious and controlled manner than is possible by the prior art method. Given a substrate that  
25 has been coated with a layer of antibody directed against the sought-after cell surface protein, the present invention facilitates the rapid assembly of a planar array of cells or decorated beads in proximity to the layer of antibodies and the subsequent disassembly  
45 of the array to leave behind only those cells or beads capable of forming a complex with the surface-bound antibody.  
30

50 A further example of interest in this category pertains to phage displays. This technique may be employed to present a layer of protein targets to bead-anchored



5 probes. Bead arrays may now be employed to identify a protein of interest. That is,  
beads are decorated with small molecule probes and an array is formed adjacent to the  
phage display. Binding will result in a probe-target complex that retains beads while  
10 others are removed when the electric field is turned off, or when light-control is applied  
5 to remove beads from the phage display. If beads are encoded, many binding tests may  
be carried out in parallel because retained beads may be individually identified  
subsequent to binding.

15 The methods of the present invention readily facilitate competitive binding  
assays. For example, subsequent to binding of a fluorescent probe to a target-decorated  
20 bead in solution and the formation of a planar bead array adjacent to the electrode,  
fluorescent areas within the array indicate the position of positive targets, and these may  
be further probed by subjecting them to competitive binding. That is, while monitoring  
the fluorescence of a selected section of the planar array, an inhibitor (for enzyme  
25 assays) or other antagonist (of known binding constant) is added to the electrochemical  
15 cell, and the decrease in fluorescence originating from the region of interest is measured  
as a function of antagonist concentration to determine a binding constant for the original  
probe. This is an example of a concatenation of analytical steps that is enabled by the  
30 methods of the present invention.

35 The fact that a probe-target complex is fixed to a colloidal bead, as in the  
20 methods of the present invention, conveys practical advantages because this facilitates  
separation of positive from negative events. Particularly when solid phase assays are  
performed on a planar substrate, an additional advantage of planar bead arrays is the  
enhancement of light collection efficiency provided by the beads, as discussed herein.

40 If desired, beads may serve strictly as delivery vehicles for small  
25 molecule probes. That is, an array of probe-decorated beads is formed adjacent to a  
target-decorated surface in accordance with the methods of the present invention.  
UV-activated cleavage of the probe from the bead support will ensure that the probe is  
45 released in close proximity to the target layer, thereby enhancing speed and efficiency of  
the assay. The identity of the particular probe interacting with the target may be  
30 ascertained from the positional location of the bead delivering the probe.

50 The methods of the present invention apply not only to colloidal beads of  
a wide variety (that need no special preparative procedures to make them magnetic, for

5 example), but also to lipid vesicles and cells that are decorated with, or contain  
embedded in their outer wall, either probe or target. The methods of the present  
invention may therefore be applied not only to bead-anchored soluble proteins but  
10 potentially to integral membrane receptors or to cell surface receptors.

5 In particular, the rapid assembly of cells in a designated area of the  
substrate surface facilitates the implementation of highly parallel cell-based functional  
assays. The present invention makes it possible to expose cells to small molecule drug  
15 candidates in solution and rapidly assemble them in the vicinity of a sensor embedded in  
the electrode surface. or to expose pre-assembled cells to such agents that are released  
20 into the adjacent liquid phase. In the simplest case, all cells will be of the same type,  
and agents will be administered sequentially. Even in this sequential version,  
electrokinetic mixing will enhance through-put. However, as described herein, the  
methods of the present invention also enable the parallel version of binding assays and  
25 thus of functional assays in a planar format by encoding the identity of different cells by  
a "Layout-Preserving Transfer" process from an 8x12 well plate, as discussed herein,  
and to isolate cells scoring positive by providing feed-back from a spatially resolved  
30 imaging or sensing process to target a specific location in the array of cells.

#### Example VIII - Screening for Drug Discovery in Planar Geometry

20 The functional elements of the present invention may be combined to  
implement procedures for handling and screening of compound and combinatorial librar-  
35 ies in a planar format. The principal requisite elements of this task are: sample and  
reagent delivery from the set of original sample reservoirs, commonly in a format of  
8x12 wells in a microtiter plate, into a planar cell; fabrication of planar arrays of targets  
40 or of probe-target complexes adjacent to the planar electrode surface prior to or  
subsequent to performing a binding assay; evaluation of the binding assay by imaging  
the spatial distribution of marker fluorescence or radioactivity, optionally followed by  
45 quantitative pharmacokinetic measurements of affinity or binding constants; isolation of  
beads scoring positive, and removal from further processing of other beads; and  
30 collection of specific beads for additional downstream analysis. The present invention  
relates to all of these elements, and the fundamental operations of the invention provide  
50 the means to concatenate these procedures in a planar format.

5 A central issue in the implementation of cost-effective strategies for  
modern therapeutic drug discovery is the design and implementation of screening assays  
in a manner facilitating high throughput while providing pharmacokinetic data as a basis  
10 to select promising drug leads from a typically vast library of compounds. That is,  
5 molecular specificity for the target, characterized by a binding constant, is an important  
factor in the evaluation of a new compound as a potential therapeutic agent. Common  
15 targets include enzymes and receptors as well as nucleic acid ligands displaying  
characteristic secondary structure.

The emerging paradigm for lead discovery in pharmaceutical and related  
10 industries such as agricultural biotechnology, is the assembly of novel synthetic  
compound libraries by a broad variety of new methods of solid state "combinatorial"  
20 synthesis. Combinatorial chemistry refers to a category of strategies for the parallel  
synthesis and testing of multiple compounds or compound mixtures in solution or on  
solid supports. For example, a combinatorial synthesis of a linear oligopeptide  
25 containing  $n$  amino acids would simultaneously create all compounds representing the  
possible sequence permutations of  $n$  amino acids. The most commonly employed  
implementation of combinatorial synthesis relies on colloidal bead supports to encode  
30 reaction steps and thus the identity of each compound. Beads preferred in current  
practice tend to be large (up to 500 microns in diameter) and porous to maximize their  
20 compound storage capacity, and they must be encoded to preserve the identity of the  
compound they carry.

Several methods of encoding, or binary encoding, of beads are available.  
Two examples are as follows. First, beads may be labeled with short oligonucleotides  
40 such as the 17-mers typically employed in hybridization experiments. The sequence of  
25 such short probes may be determined by microscale sequencing techniques such as direct  
Maxam-Gilbert sequencing or mass spectrometry. This encoding scheme is suitable  
when the task calls for screening of libraries of nucleic acid ligands or oligopeptides.  
45 Second, members of a combinatorial library may be associated with chemically inert  
molecular tags. In contrast to the previous case, these tag molecules are not sequentially  
30 linked. Instead, the sequence of reaction steps is encoded by the formal assignment of a  
binary code to individual tag molecules and their mixtures that are attached to the bead  
50 in each successive reaction step. The tags are readily identified by standard analytical

5 techniques such as gas chromatography. This general encoding strategy is currently employed in the synthesis of combinatorial libraries on colloidal beads.

10 Commercial compound libraries are large, given that even for the aforementioned 17-mer, the number of sequence permutations is  $4^{17}$ , or approximately  
5  $10^{10}$ . However, the high specificity of typical biological substrate-target interactions implies that the vast majority of compounds in the collection will be inactive for any one particular target. The task of screening is to select from this large set the few potential  
15 lead compounds displaying activity in binding or in functional assays. The principal drug discovery strategy widely applied to natural compound libraries in the  
20 pharmaceutical industry is to select individual compounds from the library at random and subject them to a series of tests. Systematic screening procedures are thus required to implement the rapid screening and scoring of an entire library of synthetic compounds, in practice often containing on the order of  $10^7$  items.

25 In current practice, compounds are first cleaved and eluted from their solid supports and are stored in microtiter plates. Further sample handling in the course of screening relies primarily on robotic pipetting and transfer between different  
30 containers, typically wells in microtiter plates. While robotic workstations represent a step in the direction of automating the process, they rely on the traditional format of microtiter plates containing 8x12 wells and sample handling by pipetting and thus  
20 represent merely an incremental operational improvement. A significant additional consideration is the need to conserve reagent and sample by reducing the spatial scale of the analytical procedures.

35 The present invention provides a set of operations to realize integrated sample handling and screening procedures for bead-based compound libraries in a planar  
40 format. This will significantly reduce time and cost due to reagent and sample volumes. The principal advantage of the methods of the present invention is that they provide a large set of fundamental operations to manipulate sets of beads in a planar format,  
45 permitting the handling of beads between stations in a multi-step analytical procedure.

50 In particular, as previously described herein, the methods of the present invention facilitate the implementation of the following pertinent procedures: transfer of samples from microtiter plates to a planar electrochemical cell; formation of heterogeneous panels of target sites adjacent to the substrate surface; solid phase binding

assays; and isolation of specific beads from an array. In addition, the fundamental operations of the present invention provide the means to concatenate these procedures on the surface of a planar electrode.

As described herein for hybridization assays, several variants are possible. That is, binding assays may be performed by allowing protein targets such as enzymes to bind to compounds on the surface of a bead, either in suspension or arranged in a planar array. The common practice of combinatorial chemistry based on large porous carrier beads accommodates the concurrent handling of smaller beads to whose outer surface compounds are anchored via inert chemical spacers. Such small beads (up to 10 microns in diameter) are readily manipulated by the methods of the present invention. Large beads are used as labeled compound storage containers.

Alternatively, binding between target and a radioactively or otherwise labeled probe may occur in solution, within microtiter plate wells, if compounds have already been cleaved from their synthesis support. In that case, probe-target complexes may be captured by complexation to encoded beads in each well, for example via the secondary antibody method of coupling the protein target to a bead-anchored antibody. Bead-captured probe-target complexes are then transferred to the planar cell for proximity analysis and further processing as illustrated in Fig. 10. As shown in Fig. 10, probe-target complexes 102 are allowed to form in solution. Antibody coated beads 104 are added to the solution, resulting in a bead anchored complex 106. The bead anchored complexes 106 are deposited onto electrode 108 from wells 110, and a planar array of bead anchored complexes is formed. When fluorescent probes 114 are used, these impart fluorescence to the bead anchored complex, facilitating detection.

The methods and apparatus of the present invention are well suited to the task of identifying a small number of positive events in a large set. The imaging of an entire array of probe-target complexes is further enhanced by proximity to an area detector, and by bead lensing action. The isolation of a small number of positive scores from the array is readily achieved, for example by applying optical tweezers, as described herein. The large remainder of the array may then be discarded. This in turn considerably reduces the complexity of applying more stringent tests, such as the determination of binding constants, because these may be restricted to the few retained beads. These tests may be directly applied, without the need for additional sample

transfer to new containers, to the samples surviving the first screening pass.

#### Example IX - Hybridization Assays in Planar Array Format

The present invention can be used to implement solid phase hybridization assays in a planar array format in a configuration related to that of a protein binding assay in which target molecules are chemically attached to colloidal beads. The methods of the present invention facilitate the formation of a planar array of different target oligonucleotides for presentation to a mixture of strands in solution. Alternatively, the array may be formed subsequent to hybridization in solution to facilitate detection and analysis of the spatial distribution of fluorescence or radioactivity in the array.

Considerable research and development is presently being invested in an effort to develop miniaturized instrumentation for DNA sample extraction and preparation including amplification, transcription, labeling and fragmentation, with subsequent analysis based on hybridization assays as well as electrophoretic separation. Hybridization assays in planar array format are being developed as a diagnostic tool for the rapid detection of specific single base pair mutations in a known segment of DNA, and for the determination of expression levels of cellular genes via analysis of the levels of corresponding mRNAs or cDNAs. Hybridization of two complementary single strands of DNA involves molecular recognition and subsequent hydrogen bond formation between corresponding nucleobases in the two opposing strands according to the rules A-T and G-C; here A, T, G and C respectively represent the four nucleobases Adenine, Thymine, Guanosine and Cytosine found in DNA; in RNA, Thymine is replaced by Uracil. The formation of double-strand, or duplex, DNA requires the pairing of two highly negatively charged strands of DNA, and the ionic strength of the buffer, along with temperature, plays a decisive role.

As previously discussed herein, two principal methods to prepare heterogeneous arrays of target strands on the surface of a planar substrate are micro-dispensing ("printing") and in-situ, spatially encoded synthesis of oligonucleotides representing all possible sequence permutations for a given total length of strand. In this context, hybridization must necessarily occur in close proximity to a planar substrate surface and this condition requires care if complications from steric hindrance and from non-specific binding of strands to the substrate are to be avoided. Non-specific

5 adsorption can be a serious problem, especially in the presence of DC electric fields  
employed in current commercial designs that rely on electrophoretic deposition to  
accelerate the kinetics of hybridization on the surface. In addition, there are the techni-  
10 cal difficulties, previously discussed herein, resulting from steric hindrance and from  
5 collective effects reflecting the crowding of probe strands near the surface.

In the context of DNA analysis, colloidal (magnetic) beads are commonly  
15 used. For example, they are employed to capture DNA in a widely used screening  
procedure to select cDNAs from clone libraries. Specifically, cDNAs are allowed to  
hybridize to sequences within long genomic DNA that is subsequently anchored to  
20 magnetic beads to extract the hybridized cDNA from the mixture.

The present invention facilitates the formation of planar arrays of  
oligonucleotide-decorated colloidal beads, either prior to or subsequent to hybridization  
of a fluorescence probe strand to the bead-anchored target strand or subsequent to  
25 hybridization in free solution and bead capture of the end-functionalized target strand.  
15 In contrast to prior art methods, the present invention does not require hybridization to  
occur in the vicinity of planar substrate surface, although this is an option if  
bead-anchored probe strands are to be delivered to substrate-anchored target strands.

30 The ability to perform hybridization either in solution, on the surface of  
individual beads, or at the substrate surface provides an unprecedented degree of  
20 flexibility. In addition, the advantages of bead arrays, as described herein, make it  
feasible to select and isolate individual beads, or groups of beads, from a larger array on  
35 the basis of the score in a hybridization assay. This isolation facilitates the  
implementation of subsequent assays on the strands of interest. The fact that beads  
remain mobile also means that beads of interest may be collected in designated holding  
40 25 areas for micro-sequencing, or may be moved to an area of substrate designated for PCR  
amplification.

45 The methods of the present invention may be used to implement a  
hybridization assay in a planar array format in one of two principal variations. All  
involve the presence of the entire repertoire of beads in the planar array or panel formed  
30 adjacent to the electrode surface for parallel read-out. As with heterogeneous panels in  
50 general, the arrangement of beads within the array is either random (with respect to  
chemical identity), and the identity of beads scoring high in the binding assay must be

5 determined subsequently, or it is spatially encoded by invoking the "Layout-Preserving Transfer" method of sample loading described herein.

10 The former variant is readily implemented and accommodates array formation either prior to or subsequent to performing the binding assay. For example, binding may be performed in suspension before beads are assembled into the array. As 5 with the aforementioned cDNA selection procedure, the method of the present invention also accommodates the use of beads as capture elements for end-functionalized target 15 DNA, for example, via biotin-streptavidin complexation. In this latter case, beads serve as a delivery vehicle to collect all probe-target complexes to the electrode surface where 20 they are assembled into an array for ease of analysis. In particular, proximity CCD detection of beads on electrodes will benefit from the lensing action of the beads in the array. This version of the assay is preferably used if only a small number of positive scores are expected.

25 Hybridization to a pre-formed bead array can take advantage of a variant of the assay which preserves spatial encoding. An array of bead clusters is formed by 15 the "Layout-Preserving Transfer" method previously described herein, and exposed to a mixture of cDNAs. The resulting spatial distribution of fluorescence intensity or 30 radioactivity reflects the relative abundance of cDNAs in the mixture. This procedure relies on the detection of a characteristic fluorescence or other signal from the 20 probe-target complex on the surface of a single bead. Given the fact that the array is readily held stationary by the methods of the present invention, image acquisition may 35 be extended to attain robust signal-to-noise for detection of low level signals. For example, a signal generated by a bead of 10 micron diameter with at most  $10^8$  probe-target complexes on the surface of the bead may be detected. Bead lensing action 40 also aids in detection. 25

45 As with the implementation of drug screening, the functional elements of the present invention may be combined to perform multiple preparative and analytical procedures on DNA.

#### 30 Example X - Alignment and Stretching of DNA in Electric Field-Induced Flow

50 The present invention can be used to position high-molecular weight DNA in its coiled configuration by invoking the fundamental operations as they apply to other



colloidal particles. However, in addition, the electrokinetic flow induced by an electric field at a patterned electrode surface may be employed to stretch out the DNA into a linear configuration in the direction of the flow.

Procedures have been recently introduced which rely on optical imaging to construct a map of cleavage sites for restriction enzymes along the contour of an elongated DNA molecule. This is generally known as a "restriction map". These procedures, which facilitate the study of the interaction of these and other proteins with DNA and may also lead to the development of techniques of DNA sequencing, depend on the ability to stretch and align DNA on a planar substrate.

For individual DNA molecules, this has been previously achieved by subjecting the molecule to elongational forces such as those exerted by fluid flow, magnetic fields acting on DNA-anchored magnetic beads or capillary forces. For example, DNA "combs" have been produced by simply placing DNA molecules into an evaporating droplet of electrolyte. If provisions are made to promote the chemical attachment of one end of the molecule to the surface, the DNA chain is stretched out as the receding line of contact between the shrinking droplet and the surface passes over the tethered molecules. This leaves behind dry DNA molecules that are attached in random positions within the substrate area initially covered by the droplet, stretched out to varying degrees and generally aligned in a pattern of radial symmetry reflecting the droplet shape. Linear "brushes", composed of a set of DNA molecules chemically tethered by one end to a common line of anchoring points, have also been previously made by aligning and stretching DNA molecules by dielectrophoresis in AC electric fields applied between two metal electrodes previously evaporated onto the substrate.

The present invention invokes electrokinetic flow adjacent to an electrode patterned by UV-mediated regrowth of oxide to provide a novel approach to the placement of DNA molecules in a predetermined arrangement on a planar electrode surface, and to the stretching of the molecules from their native coil configuration into a stretched, linear configuration that is aligned in a pre-determined direction. This process is shown in Fig. 11 and is accomplished by creating controlled gradients in the flow vicinity across the dimension of the DNA coil. The velocity gradient causes different portions of the coil to move at different velocities thereby stretching out the coil. By maintaining a stagnation point at zero velocity, the stretched coil will be fixed

5 in position. This method has several advantages over the prior art approaches. First,  
DNA molecules in their coiled state are subjected to light control to form arrays of  
desired shape in any position on the surface. This is possible because large DNA from  
10 cosmids or YACs forms coils with a radius in the range of one micron, and thus acts in  
5 a manner analogous to colloidal beads. A set of DNA molecules may thus be steered  
into a desired initial arrangement. Second, UV-patterning ensures that the elongational  
force created by the electrokinetic flow is directed in a predetermined direction. The  
15 presence of metal electrodes in contact with the sample, a disadvantage of the  
dielectrophoretic prior art method, is avoided by eliminating this source of  
contamination that is difficult to control especially in the presence of an electric field.  
20 On patterned Si/SiO<sub>x</sub> electrodes, flow velocities in the range of several microns/second  
have been generated, as required for the elongation of single DNA molecules in flow.  
Thus, gradients in the flow field determines both the fractional elongation and the  
orientation of the emerging linear configuration. Third, the present invention facilitates  
25 direct, real-time control of the velocity of the electric field-induced flow, and this in turn  
conveys explicit control over the fractional elongation.

30 This invention is also for a method and apparatus to direct the lateral  
motion and induce the assembly into planar arrays of cells on semiconductor surfaces in  
response to temporally and spatially varying electric fields and to projected patterns of  
20 illumination.

35 The response of different types of cells to variations in the frequency and  
voltage of the applied electric field and to specific patterns and changing intensity of  
illumination enables the on-cue assembly of multiple arrays of cells in any desired  
position on the substrate, and further enables the repositioning, disassembly and re-  
40 assembly and more generally the reconfiguration and segmentation of such arrays. Time-  
constant and time-varying illumination patterns are generated by an optically  
programmable illumination pattern generator may be used and can be based on a  
45 graphical representation of the pattern created on a personal computer. The  
novel methodology of image cytometry introduced in the present invention relates to the  
30 general fields of molecular and cellular biology and has applications in the fields of  
diagnostics, genetic analysis, cell biology and drug discovery. The image cytometry  
50 facilitates a multiplicity of cell-based biological and biochemical procedures based on the

quantitative analysis of images recorded from array of cells. As with Random Encoded Array Detection (READ) or Programmable Array Reconfiguration and Segmentation (PARSE), these assays are performed in a highly parallel format in a small volume of sample on the surface of a semiconductor wafer.

The novel method of image cytometry according to the present invention may be used in a number of different applications, including:

- cell-typing analysis based on the decoration of cell-surface markers with tagged probes such as fluorescence labeled antibodies or antibodies that are coupled to fluorescent beads. Specific applications include bacterial strain typing; HLA typing (transplantation donor-recipient matching); immunophenotyping;
- diagnostic assays based on the identification of cell types and quantitative evaluation of relative counts of different cell types in a given sample, for example, the determination of the fraction of CD-4 cells in a population to assess levels of HIV infection;
- on-chip fractionation of a mixture of cells into sub-populations based on:
  - selective affinity of cell phenotypes using antibodies or lectin (Nordon et al, "An Experimental Model of Affinity Cell Separation", Cytometry 16, 25-33 (1994), the contents of which are incorporated herein by reference; see also: "panning" for clones expressing specific antibodies). Using beads displaying such antibodies or lectins, cross-linking between cells may be induced to form clusters that are readily distinguished and separated from individual cells;
  - differential mobility in response to time-varying or to time-constant but spatially varying (inhomogeneous) electric fields or to gradients in electrochemical environment created by LEAPS near an electrolyte-insulator-semiconductor (EIS) interface;
  - combinations with controlled flow configurations such as those in various realizations of field flow fractionation (for example, see: Bigelow et al, "Separation of B and T lymphocytes by a hybrid field-flow fractionation/adhesion chromatography technique", J. Immunological Methods 117, 289-293 (1989); Markx et al, "Dielectrophoretic characterization and separation of microorganisms", Microbiology 140, 585-591 (1994); Huang et al, "Introducing Dielectrophoresis as a New Force Field for Field Flow Fractionation", Biophysical J. 73, 1118-1129 (1997); the contents of all of which are incorporated herein by reference);

- 5 - cell-based functional assays monitoring the levels of expressed cell-surface  
markers or the activation or regulation of pathways, such as signal transduction  
pathways and metabolic pathways. The expression of the surface markers may be  
10 detected by affinity binding between the markers and fluorescence or enzyme labeled  
5 probes. The endpoints of the pathways may be analyzed through intracellular fluorescent  
signal generated by enzymes such as luciferin (PE Corporation/Tropix) or  $\beta$ -lactamase  
(Aurora);
- 15 - cell secretion-based analysis that could be used in drug and ligand screening;  
- cell-cell interactions revealed by adhesion markers on cell surfaces;  
10 - cytotoxicity assays based on the labeling of indicators such as  
20 phosphatidylserine which appears on the outer membrane of cells in the early stages of  
apoptosis; and
- chemotaxis analysis based on the tracking of cells positioned in controlled  
25 configurations in known spatial relation to gradients in the concentration of chemotactic  
15 agents.

The image cytometry according to the present invention permits the  
efficient realization of a wide variety of cell-based functional as well as diagnostic assays  
30 based on the analysis of multi-color fluorescence or other optical markers imaged on the  
surfaces of individual cells. In addition, the methodology also permits the separation and  
20 fractionation of multiple cell types in a mixture based on differential response to  
35 electro-optic stimuli as disclosed herein. Imaging and profiling applications may invoke  
a variation of the READ format while fractionation and sorting into multiple sub-  
populations may invoke the PARSE format.

40 The integration of this novel methodology of cellular analysis with sample  
25 preparation (electric-field induced lysis of cells) along with the technology of optically  
programmable bead arrays as disclosed herein creates a novel platform that combines an  
"on-chip" format of quantitative molecular interaction analysis and cellular analysis with  
45 microfluidics and optical imaging into a novel, multi-purpose assay platform.

*Imaging of cells* - In its simple form, image cytometry is practiced in the  
30 doctor's office when blood or other liquid samples are spotted onto segmented glass  
slides and populations of cells are "manually" counted. These and related simple  
50 implementations of cellular analysis invoke the simple spotting of cells onto solid

5 supports where cells settle under gravity, typically in random positions at low lateral  
density. However, even in this simple format, certain advantages of the approach have  
been recognized (Galbraith et al, "Imaging Cytometry by Multiparameter  
10 Fluorescence", Cytometry 12, 579-596 (1991), the contents of which are incorporated  
5 herein by reference): the power of image analysis can be brought to bear to facilitate the  
analysis of multi-color fluorescence images. A major drawback is the very low  
15 "throughput" of this approach, given the low density of cells which must be maintained  
to avoid the risk of forming three-dimensional clusters in which the view of some of the  
cells would be obstructed. A further drawback is that methods of cellular analysis  
20 invoking differential mobility of cells in response to external stimuli are not available.

*Flow cytometry* - Flow cytometry and its application to fluorescence-  
activated cell sorting rely on a serial mode of analysis that requires the capture of  
individual cells into droplets which are passed, one at a time, through a small  
25 observation window. During a short time, e.g., 1-10 $\mu$ s, all measurements on a given  
15 cell must be completed. For example, the simultaneous analysis of a small number of  
cell surface markers represents the state of the art in flow cytometry (Beavis and  
Pennline, "Detection of Cell-Surface Antigens Using Antibody-conjugated  
30 Fluorospheres", BioTechniques 21, 498-503 (1996), the contents of which are  
incorporated herein by reference). Flow cytometric analysis represents just the last step  
20 in an assay: the instrumentation does not lend itself to the integration of liquid handling  
and automation of bioanalytical assays. This is in significant contrast to the present  
35 invention which integrates the optical manipulation of bead arrays and cellular assembly  
technology with microfluidics and optical imaging into a miniaturized platform for  
quantitative molecular and cellular analysis.

40 25 Recent developments of smaller and more specialized instruments  
notwithstanding, the intrinsic complexity of flow cytometry requires specialized and  
expensive instrumentation including multiple laser sources of high power, matching  
45 detectors and fast electronics to handle the acquisition and processing of signals during  
the short available interrogation interval during which each cell traverses the detection  
30 window. Sorting requires additional steps and equipment. Even after many years of  
commercial development, the operation of flow cytometers requires trained personnel to  
50 handle calibration and ensure reliable performance. The cost of commercial instruments,

for example, those offered by Becton-Dickinson, Beckman Coulter or Cytomation is prohibitive for all but major medical centers or research institutions.

*Sorting and separation of cells by affinity and/or flow* - Several realizations have been described for the fractionation of cells by selective affinity for, and hence differential retention by a nearby solid phase or by differential mobility in superimposed force fields such as a fluid flow with superimposed transverse electric field (field flow fractionation or FFF) or by combinations thereof. Differential dielectrophoretic levitation of cells above a set of interdigitated electrodes and fractionation in a transverse flow represents a recent addition to this set of techniques which offers the potential for differential blood cell analysis. As with other FFF techniques, the latter method relies on downstream detection of individual cells and thus resembles flow cytometry with respect to the implied serial processing to establish the identity of sub-populations. A further disadvantage is the requirement for an elongated channel to permit the spatial separation between sub-populations.

#### *Cytometry According to the Present Invention*

Cytometry according to the present invention subsumes the entire set of capabilities for cellular analysis illustrated above into a common platform which is miniaturized and integrated with microfluidics and optical imaging and compatible with quantitative bead array format of molecular interaction analysis whose capabilities may be used in "multiplexed" fluorescence immunoassays and DNA hybridization assays.

The specific advantages of the cytometry according to the present invention include:

- high throughput resulting from the high densities of the cellular assemblies (for example,  $10^4$  -  $10^6/\text{mm}^2$ ) and the parallel mode of signal acquisition afforded by imaging;
- integration of sample preparation, fluid handling and assay readout, for example, integration with on-chip assay formats;
- parallel processing based on imaging of multiple colors and morphological parameters;
- light-directed transport and assembly of cells;
- light-guided separation and fractionation of cells into sub-populations based on optically programmable illumination patterns and frequency dispersion in the

5 dielectrophoretic response; and

- significant reduction in expense of instrumentation.

The method and apparatus of the present invention enables the parallel  
10 processing of large numbers of cells assembled into dense planar assemblies; typically,

5  $10^4$  cells of  $10\mu\text{m}$  diameter will be accommodated in an area as small as  $1\text{mm}^2$ .

Smaller bacterial cells such as *E.coli* can be accommodated in even higher density. In  
15 significant contrast to flow cytometry, signals are collected simultaneously from a large  
number of cells by way of imaging, and the rate of acquiring signal is very high even  
when cells are held stationary.

10 The present invention presents a very significant advantage, especially  
when multiple colors are to be discriminated and are to be quantitatively determined by  
20 analysis of multi-color images of individual cell surfaces. Thus, the cytometry  
according to the present invention will readily accommodate multiple cell surface  
25 markers and additional morphological indicators for individual cells (size, shape) or  
mutually interacting clusters of cells. This is generally possible even without specialized  
15 equipment. In fluorescence immunoassays using the READ bead array format, the  
capability has been demonstrated using an inexpensive consumer-grade video CCD  
30 camera, of detecting signals from individual beads displaying only 1000 labeled antibody  
molecules. Higher levels of sensitivity are possible for detection of markers on cell  
20 surfaces because fluorescent beads may be used as the label to decorate the cells.

#### 35 A. Cells form arrays in response to AC electric fields

Referring now to Figure 12, therein is illustrated *E.coli* cells (grown to  
40 log phase and suspended in 280 mM mannitol solution at the density of  $10^8/\text{ml}$ ) which  
25 form arrays under electric field. On the left side of the field, the cells are assembled at  
very high density. Some singlet cells at the right side are moving to the left to join the  
array. (Bright field image. 25x obj, NA 0.45. Electric field: 5V, 200 Hz).

45 Referring now to Figure 13, therein is illustrated the same effect as in  
Figure 12, with *S. cerevisiae* cells (baker's yeast). (Bright field image. 25x obj, NA  
30 0.45; electric field: 5V, 400 Hz).

#### 50 B. Cells assemble in accordance with projected illumination patterns.

Referring now to Figure 14, therein is illustrated *E. coli* forming diamond shapes that are projected to the surface by an LCD panel controlled by appropriate graphics software; (20x objective, NA 0.28; Electric field: 10V, 7 kHz).

Referring now to Figure 15, therein is illustrated *S. cerevisiae* responding to two circular illumination patterns; (20x objective, NA 0.28; Electric field: 15 V, 15 kHz).

### C. Light-directed cell transport and reconfiguration of cellular assemblies

By enabling the controlled transport of cells and cellular assemblies in accordance with the PARSE format, the cytometry according to the present invention provides an entire additional level of cellular analysis which has the inherent capability to fractionate cell populations "in-situ".

The principal approach according to the present invention is to generate gradients in electrochemical properties of the interface to which sub-populations of cells exhibit a differential response. Such gradients may be produced by invoking patterns of illumination in geometry, as disclosed herein. Examples in the figures illustrate that cells respond to these patterns in a variety of ways.

For example, cells may be collected into illuminated regions (Fig. 16; *S. cerevisiae* responding to the illumination. Arrow indicate the direction cells are moving in. (20x objective, NA 0.28; Electric field: 15 V, 15 kHz)) or expelled from illuminated regions (Fig. 17: *E. Coli* cells being repelled from the illumination circle. (20x objective, NA 0.28; Electric field: 10 V, 7 kHz; low substrate doping level (1 Ohm cm-10 Ohm cm)); "channeled" by reconfiguring illuminated regions or "dragged" and "dropped" by displacing illuminated regions (Fig. 18: *E. coli* cells following the expansion of the rectangular illuminated shape (20x objective, NA 0.28; Electric field: 15 V, 8 kHz)).

Cells also may be "trapped" in well defined regions in response to light-induced spatial gradients in the electrical properties of the EIS structure (Fig. 19; *E. coli* cells being trapped by the light stripe at the left and being prevented from moving to the circular region on the right. (20x objective, NA 0.28; Electric field: 15 V, 8 kHz)).

This set of fundamental operations permits the creation of spatially varying electrochemical potentials and corresponding lateral, highly inhomogeneous electric fields. In addition, the cytometry according to the present invention permits the



programmable lateral displacement and reconfiguration of illumination patterns so that the location and the strength of the spatial inhomogeneities in electric fields can be adjusted at will.

In addition, the cytometry according to the present invention also provides frequency-dependent forces to which different constituents within mixtures of cells will generally exhibit differential responses. An example of segregation between different types of cells is shown in Fig.20 As shown in Figure 20, when an illuminated region containing both types of cells is suddenly expanded or "dragged", *E. coli* cells (left) respond faster than the larger *S. cerevisiae* cells (right), resulting in segregation. (Bright field image. 25x obj, NA 0.45 Electric field: 5V, 400 Hz).

Specifically, on the basis of the known dielectrophoretic properties of various cells, fractionation will be possible on the basis of characteristic frequency dispersion. For example, the characteristic transition of cells from positive to negative dielectrophoresis (as evidenced in electrorotation; see e.g.: Huang et al, op. cit.) will be the basis for the fractionation of cell mixtures in light-induced gradients in electrochemical EIS potential (and the resulting inhomogeneous fields): different cells will be expelled from confined regions on the substrate at characteristic frequencies and light intensities.

#### D. Non-viable cells behave differently than the viable cells.

The present inventors have observed that heat treated *S. cerevisiae* cells stop responding to the electric field and light after the first ten to twenty seconds and adhere to the substrate.

**Example: Immunophenotyping** - In a manner analogous to READ, a cellular array format will provide a sensitive and highly parallel mode of monitoring the presence of characteristic cell surface markers as illustrated in Figure 21. For example, groups of lymphocyte cells can be put into array format on the substrate. Antibodies for the cell surface markers (CD3, CD4, CD8, CD56, etc.) are coupled to small microspheres that are differentially coded. The mixture of beads with different antibodies are directed to the cells arrays using the optical arrangement principles of the present invention. Antibodies on the bead surface will bind to the cell surface markers, thus, the correspondent beads will link to the cells. The number of surface markers which can be detected simultaneously depends on chemical coding capacity of the bead. Hundreds of

chemical codings may possibly be used. Signal integration capability according to the present invention enables more accurate and quantitative analysis of the cells.

*Example: Multiplexed Affinity Detection* - A novel format enabling the simultaneous affinity identification of multiple types of cells is shown in Fig. 22. A random encoded bead array of antibodies will provide a novel, highly diverse set of anchoring points for lymphocytes displaying cognate cell surface antigens. Under conditions of controlled stringency, flow-induced shear will remove cells that are not specifically bound while leaving "positives" attached to individual beads. Focusing on this downstream step takes advantage of the intrinsic "concentration" of sample on the cell surface and the ability to capture, with a single antibody probe, multiple pathogens sharing common processed antigenic determinants.

*Example: Light-guided Sorting and Isolation* - Combination of AC electric fields with light-controlled modulation of surface-electrochemical properties is particularly powerful. The illuminated areas mimics the behavior of patterned electrodes and can be repositioned in real time to isolate designated cells. Cells can be sorted and isolated due to their natural properties or due to the property change caused by specific binding to probe labeled beads. The model is illustrated in Fig.23. Cells can be recovered after the isolation and further culturing or analysis may be performed. Specific lymphocyte cells and cancer cells may be of particular interest for such cell sorting and analysis.

*Example: Cell-based Functional Assays* - Cell-based functional assays may be performed in a highly efficient manner by assembling cellular arrays and imaging reaction of the cells as shown in Fig. 24. The reaction may be the direct change of extracellular markers, or the endpoints of some metabolic pathways. The endpoints may be intracellular fluorescence signal generated by enzymes such as  $\beta$ -lactamase whose substrate can penetrate the cells, thus, the signal may be directly quantitatively detected by imaging (Zlokarnik et al., "Quantitation of Transcription and Clonal Selection of Single Living Cells with  $\beta$ -lactamase as Reporter", Science 279, 84-88 (1998); see also: Bronstein et al, "Chemiluminescent and Bioluminescent Reporter Gene Assays", Anal. Biochemistry 219, 169-181 (1994); the contents of all of which are incorporated herein

5 by reference). If combined with microfluidic control, such cell-based functional assays have tremendous applications in drug screening and ligand analysis. Cells may be placed at the same location as the chemical carrier beads. Under temporal and spacial controlled  
10 release of the compounds or ligands, response of the cells may be recorded. After image analysis, compounds or ligands that generate the desired cell response may be easily identified. Because the cells may be directly imaged, cell lysis and the following assays will generally not be necessary as part of the traditional cell-based analysis.  
15 Furthermore, the controlled assembly of the cells and chemical carrying beads, according to the present invention, may eliminate the use of microtiter plates in the cell-based screening.  
20

*Example: Integrated Format Profiling of Secreted Cellular Markers* - Control of cell assembly and microfluidic pathway enables direct in situ screening of secreted cellular markers. This model is illustrated in Fig. 25. Cellular arrays are assembled in one  
25 compartment of a two-compartment liquid reactor and are exposed to candidate drugs; the response is measured by monitoring a panel of secreted markers such as cytokines. Cell secretion are collected and introduced into a second compartment where they are profiled based on the response of a pre-deposited bead array using the READ format. Such a two-compartment reactor may be operated in parallel for multiple drug screening  
30 or kinetic analysis.  
35

*Example: Cell-to-Cell Interaction* - Cell-to-cell in vivo interaction was seldom studied directly due to the lack of control in cell manipulation. The present invention provides a powerful tool in this area. As illustrated in Fig. 26, cells may be temporally and  
40 spatially controlled to favor the binding between the cells. Alternatively, cells may be directed to preassembled cell arrays for cell binding analysis. It may also be possible to study the interaction between cells such as T cells, B cells and polymorphonuclear leukocytes, in the activation of immune responses.  
45

30 *Example: Chemotaxis Analysis* - Cells can be positioned in a controlled configuration to test their chemotaxis properties. This model is illustrated in Fig. 27. For example, *D. discoideum* cells may be tested under cAMP or specific polypeptide gradient. (Parent &  
50

5 Devreotes, "A cell's sense of direction", Science 284, 765-770 (1999); the contents of which are incorporated herein by reference). Response of the receptors, G proteins, actin or actin-binding proteins may be detected through fluorescence signal generated by  
10 their correspondent fusion protein with GFP (Green Fluorescent Protein). Movement of  
5 the cells may also be recorded. More importantly, the force of the movement may be measured by comparing to the energy used in the light control of the cells.

15 The present invention may be used for cell identification and detection, for example, in immunophenotyping and affinity cell detection. In this application, affinity probes are coupled to beads on a solid surface. The beads are chemically and  
10 spatially coded. The cells are transported to the desired position using light control.  
20 Cells with the surface markers will bind at the correspondent positions. The advantages of this approach include a simpler and less expensive system, a higher degree of throughput for parallel immunophenotyping, the manipulation of a large number of cells, thus the detection of rare events, and better integration between the reaction and the  
25 detection steps, thus allowing the entire process to be performed on-chip.

30 The present invention may also be used in cell sorting, fractionation and isolation. In this case, cells can be separated according to their intrinsic properties, such as their dielectrophoretic mobility, viability, size, etc. Cells can also be sorted by their association with antibody coupled beads. The advantages of this approach include  
20 multiple cell sorting, simpler substrate fabrication, and flexible and programmable control.

35 The present invention may also be used for screening, for example, cell mediated drug and ligand screening, or cell secretion mediated screening. In this approach, cells may be transported and assembled at the desired position. Chemical  
40 compounds or ligands are released at the corresponding position. The response of the cells may then be observed through their reporter genes. The advantages of this approach include higher throughput, a simpler system without the complication of  
45 automated pipetting machines, and the possible combination with microscopy for simpler and more direct assays.

50 The present invention may also be used for cell to cell interaction, for example, in suspension or on surface interaction. In this approach, cells may link with each other through their surface interaction. Movement of the cells may then be guided

5

by light. The advantages of this approach include the ability to study the interaction between cells.

10

While the invention has been particularly shown and described with reference to a preferred embodiment thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the spirit and scope of the invention.

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